

Catalog Number AP 014 NW V10

## Highlights:

- Test corn, cotton, canola or soybean
- High Sensitivity Protocol detects the PAT enzyme from the *pat* gene (PAT/*pat*) found in 0.5% T25 corn
- Rapid Protocol screens individual seeds or leaf samples for the presence of PAT/*pat*

## Contents of Kit:

- 10 antibody-coated 96-well plates
- PAT/*pat* Enzyme Conjugate
- Substrate

## Intended Use

The QualiPlate Kit for LibertyLink PAT/*pat* is designed for the qualitative laboratory detection of phosphinothricin acetyl transferase enzyme (PAT) coded for by the *pat* gene in grain, leaf, or seed. Two assay protocols are presented. The High Sensitivity Protocol will detect the PAT enzyme from the *pat* gene (PAT/*pat*) found in 0.5% LibertyLink T25 corn, 40% Bt11(T14) corn, 20% Herculex™ I corn, 5% WideStrike™ cotton, 0.5% T45 LibertyLink canola, and 1.0% LibertyLink soybean, and requires 2.5 hours to run. The Rapid Protocol (one hour total) is intended for use in screening individual seeds or leaf samples for the presence of PAT/*pat*.

NOTE: LibertyLink PAT from the *bar* gene, at concentrations present in StarLink® corn, LL25 cotton, or InVigor™ canola, is not detected in either format.

## How the Test Works

This QualiPlate Kit is a “sandwich” Enzyme-Linked ImmunoSorbent Assay (ELISA). In the test, **corn, cotton, canola, or soybean** sample extracts are added to test wells coated with antibodies raised against PAT from the *pat* gene. Any residues present in the sample extract bind to the antibodies, and are then detected by addition of enzyme (horseradish peroxidase)-labeled PAT/*pat* antibody.

After a simple wash step, the results of the assay are visualized with a color development step; color development is proportional to PAT/*pat* concentration in the sample extract.

*Lighter color = Lower concentration*

*Darker color = Higher concentration*

## Materials not Provided

- PBS/0.05% Tween-20 Wash Buffer (may be purchased in 1L dry packets from Sigma Chemicals, Cat#P-3563, or prepared from salts on site). Store at controlled ambient temperature for up to one week, then discard.
- 1 N Hydrochloric acid (HCl) Stop Solution. Prepare by adding 83 mL of concentrated HCl (36.5-38.0%) to 917 mL of distilled or deionized water; work in a fume hood and use proper protective gear. This reagent may be stored at room temperature for 2 years.
- distilled or deionized water for preparing Wash/Extraction Buffer
- glass bottles or flask plus graduated cylinder with 1 liter capacity for preparation and storage of Wash/Extraction Buffer
- test or centrifuge tubes for extraction of grain
- Tissue Extraction Kit: snap-cap tubes and pestles for extraction of leaf samples (EnviroLogix Cat. No. ACC 002, 100/package)
- centrifuge capable of 5000 x g
- disposable tip, adjustable air-displacement pipettes which will measure 50 and 100 microliters (µL)
- marking pen (indelible)
- tape or Parafilm®
- timer
- microtiter ELISA plate reader



Prepare Wash/Extraction Buffer

**Wash/Extraction Buffer:**

Order item # P-3563 from Sigma Chemical Co. (St. Louis, MO), or prepare the equivalent. Store refrigerated when not in use; allow to come to room temperature prior to assay. Use this buffer for the wash step of the assay, and to extract all corn samples.

**Stop Solution:** 1 N Hydrochloric acid (HCl). Prepare by adding 83 mL of concentrated HCl (36.5-38.0%) to 917 mL of distilled or deionized water; work in a fume hood and use proper protective gear. This reagent may be stored at room temperature for 2 years.

- wash bottle, or microtiter plate or strip washer
- multi-channel pipette that will measure 50 and 100  $\mu$ L
- racked dilution tubes for loading samples into the plate with a multi-channel pipette, or the equivalent
- orbital plate shaker (optional)

## Choose the Assay Protocol

Choose the assay protocol (High Sensitivity or Rapid) that best suits the samples to be tested according to the table below. A “yes” in a column indicates that the indicated trait can be reliably detected in the assay protocol named. PAT/*pat* protein is expressed at levels too low to be reliably detected in the crops/tissues with a “no” statement.

Limits of detection (LOD) stated for bulk seed are in units of % by weight of PAT/*pat*-expressing seed in a bulk seed sample.

Crop	Event or Tradename	Leaf Rapid Protocol	Single Seed Rapid Protocol	Bulk ground seed High Sensitivity Protocol
Corn	LibertyLink T25	Yes	Yes	0.5% LOD
Corn	Herculex I	Yes	No	20% LOD
Corn	Bt11/T14	No	No	40% LOD
Cotton	WideStrike	No	No	5% LOD
Canola	LibertyLink T45	Yes	Yes	0.5% LOD
Soybean	LibertyLink	Yes	Yes	1.0% LOD

## Sample Preparation

Note: PAT/*pat* protein is not stable in solution. Extraction should be performed in 30 minutes or less. Do not extract overnight.

### High Sensitivity Protocol for Ground Grain/Seed

**Corn:** This protocol requires that a small sample be analyzed. It is essential that this sample be well mixed and representative of the larger bulk. The test will detect 0.5% T25, 40% Bt11, or 20% Herculex I corn containing PAT/*pat*.

**Note:** Thorough mixing of the bulk grain sample and determination of an appropriate sampling plan are critical to the results of this testing, and are the responsibility of the user of this test kit. The USDA/GIPSA has prepared several guidance documents to address the issues involved in obtaining representative grain samples from static lots - such as trucks, barges, and railcars - and for taking samples from grain streams. Sampling plans should be chosen that best meet the needs of both the buyer and seller in terms of acceptable risks. Increasing the number of kernels in the sample and taking multiple samples will increase the likelihood of obtaining representative samples, and maximize the probability of detecting any contamination in the grain lot. For further information on USDA/GIPSA guidelines for obtaining representative samples and assessing detection probabilities for biotech grain, see the websites listed on page 3.

It is the responsibility of the user to ensure proper sampling and thorough mixing prior to analysis. Once representative samples have been obtained from the truck or container, they can be reduced in size using a splitter and uniformly ground and mixed.

**The finer the grind, the faster and more efficient the extraction.**

1. Weigh 5 grams of ground corn sample into a 30+ mL capacity vial or tube.
2. Add 12.5 mL of Wash/Extraction Buffer to each 5 gram sample. For all other grain sample sizes, add Wash/Extraction Buffer at the rate of 2.5 mL per gram of

**Reference Websites:**

<http://archive.gipsa.usda.gov/reference-library/handbooks/grain-insp/grbook1/bk1.pdf>

USDA *Grain Inspection Handbook, Book 1, Grain Sampling*. This document provides a comprehensive overview of recommended sampling guidelines for static lots and grain streams. It reviews the various types of equipment and strategies that can be used to obtain a representative grain sample from different types of containers.

<http://archive.gipsa.usda.gov/biotech/sample2.htm>

Guidance document entitled *Sampling for the Detection of Biotech Grains*, which provides important statistical sampling considerations when testing for the presence of biotech grains. It covers the basis for making probability determinations in accepting lots based upon different assumptions with respect to sample size, number of samples, sample preparation, etc.

<http://archive.gipsa.usda.gov/biotech/sample1.htm>

*Practical Application of Sampling for the Detection of Biotech Grains*. This one-page application guide provides a table that gives sample sizes for selected lot concentrations and probability of rejecting the specified concentrations. It also provides a formula for making the calculation for other combinations.

<http://archive.gipsa.usda.gov/biotech/samplingplan1.xls>

This website provides a simple to use Sample Planner (29k Excel Spreadsheet). The planner allows you to enter different assumptions in terms of sample size, number of samples, acceptable quality level and to determine the probability of accepting lots with given concentration levels. It also plots the probabilities in graph form for easy interpretation. Specific data can be saved for documentation and future analyses.

grain. Cap and shake vigorously by hand or vortex for 20-30 seconds. Let stand at room temperature for 10 to 30 minutes to extract.

3. The extracted samples/controls must be clarified by centrifuging the extract at 5000 x g for 5 minutes. Insert a pipette tip below any floating lipid layer and above the pellet to remove the clarified sample.

**Canola:**

1. Weigh out 2 grams of canola sample into a mesh bag (EnviroLogix Cat No. ACC 021). Use a rubber mallet to crush the seeds thoroughly. After crushing, run your fingers over the bag to make sure there are NO uncrushed seeds remaining.
2. Add 6.0 mL of Wash/Extraction Buffer to each 2 gram sample. Thoroughly massage the extraction buffer around in the bag until the entire sample is wet. Allow the sample to extract for 10 to 30 minutes at room temperature.
3. Remix the sample using your finger on the outside of the bag until you feel the sample is homogenous. Tilt the bag slightly and use your fingers to pull the extraction buffer over to one corner. Pipette out a sample (the sample will contain particles).
4. The extracted samples must be clarified by centrifuging the extract at 5000 x g for 5 minutes. Insert a pipette tip below any floating lipid layer and above the pellet to remove the clarified sample.

**Bulk Cottonseed:**

1. Weigh out 5 grams ground cotton into 30+ mL capacity vial or tube.
2. Add 20 mL of Wash/Extraction Buffer to each 5 gram cotton sample. For all other cotton sample sizes, add Wash/Extraction Buffer at the rate of 4.0 mL per gram of cotton. Cap and shake vigorously by hand or vortex for 20-30 seconds. Let stand at room temperature for 10 to 30 minutes to extract.
3. The extracted samples must be clarified by centrifuging the extract at 5000 x g for 5 minutes. Insert a pipette tip below any floating lipid layer and above the pellet to remove the clarified sample.

**Soybean Grain:**

1. Weigh out 5 grams ground soybean into 30+ mL capacity vial or tube.
2. Add 25 mL of Wash/Extraction Buffer to each 5 gram soy sample. For all other soy sample sizes, add Wash/Extraction Buffer at the rate of 5.0 mL per gram of soy. Cap and shake vigorously by hand or vortex for 20-30 seconds. Let stand at room temperature for 10 to 30 minutes to extract.
3. The extracted samples must be clarified by centrifuging the extract at 5000 x g for 5 minutes. Insert a pipette tip below any floating lipid layer and above the pellet to remove the clarified sample.

**Rapid Protocol for Screening Single Seeds or Leaf Punches:**

**Individual seeds:**

1. Crush seeds: Corn kernels, cotton seeds, or single soybeans may be placed in a plastic bag and smashed with a hammer, then transferred to a tube; or a seed crusher/48-well plate combination may be used (Hypure #HSC-100, PerkinElmer, with Costar plate #3548, Corning Life Sciences, or equivalent). Check to be sure that all seeds have been crushed. Take extreme care not to cross-contaminate between seed samples. Canola seeds can be crushed by placing them into the wells of an uncoated 96-well plate and mashing each seed with the flat end of the pestle (EnviroLogix Cat No. ACC 002). Wipe the pestle clean on a damp paper towel between each seed.



Allow all reagents to reach room temperature before beginning



Leaf punch



Extract sample



Centrifuge to clarify sample extract  
(High Sensitivity protocol only)

2. Add 0.5 mL of Wash/Extraction Buffer to each crushed corn seed, 1 mL to each crushed cotton seed, 0.2 mL to each crushed canola seed, or 1 mL to each crushed soybean. Mix for at least 30 seconds, then allow particles to settle.

#### Leaf samples:

1. Take a single leaf punch of approximately 10 mm diameter, using a micro-tube cap. Mash the leaf tissue with a pestle matched to the micro-tube (EnviroLogix Tissue Extraction Kit).
2. Add 0.5 mL of Wash/Extraction Buffer per leaf punch. Mix for at least 30 seconds, then allow particles to settle. Take extreme care not to cross-contaminate between leaf samples.

Note: Leaf samples must be run in the assay on the same day they are punched.

## How to Run the Assay

- Read all of these instructions before running the kit.
- Allow all reagents to reach room temperature before beginning (at least 30 minutes with un-boxed plates and reagents at room temperature - do not remove plate from bag with desiccant until it has warmed up).
- Organize all sample extracts, and pipettes so that Step 1 can be performed in 15 minutes or less. The use of a multi-channel pipette is strongly recommended.
- Use a disposable-tip air-displacement pipette and a clean pipette tip to add each Calibrator and sample extract to the wells. Conjugate, Substrate, and Stop Solution may be added in the same manner; alternatively, use a repeating pipette with a disposable tip on the end of the Combitip for these three reagents.
- Use the well identification markings on the plate frame to guide you when adding the samples and reagents. For this qualitative assay, duplicate wells of the Wash/Extraction Buffer Blank (BL), user-supplied known negative sample (Negative Control, NC), and user-supplied known positive sample (Positive Control, PC), along with 90 sample extracts (S) in single wells may be run on one plate. (See the Qualitative Assay Example Plate Layout - Figure 1A, page 6).

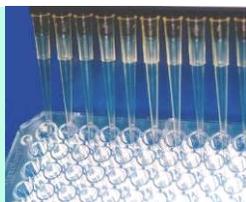
1. Add **50 µL of Enzyme Conjugate** to each well, followed immediately by **50 µL of Wash/Extraction Buffer Blank (BL)**, **50 µL of clarified Positive and Negative Control sample extracts (PC, NC)**, and **50 µL of each clarified sample extract (S)** to their respective wells, as shown in the Example Plate Layout (Figure 1A)

**NOTE:** In order to minimize setup time it is recommended that a multi-channel pipette be used in steps 1, 5, and 7.

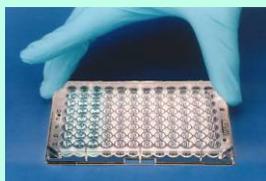
2. Thoroughly mix the contents of the wells by moving the plate in a rapid circular motion on the benchtop for a full 20-30 seconds. Be careful not to spill the contents!
3. Cover the wells with tape or Parafilm to prevent evaporation and incubate at ambient temperature according to the table below. If an orbital plate shaker is available, shake plate at 200 rpm.

<b>HIGH SENSITIVITY PROTOCOL</b>	<b>2 hours</b>
<b>RAPID PROTOCOL</b>	<b>45 minutes</b>

4. After incubation, carefully remove the covering and vigorously shake the contents of the wells into a sink or other suitable container. Flood the wells completely with Wash/Extraction Buffer, then shake to empty. Repeat this wash



*Add Conjugate, controls and samples*



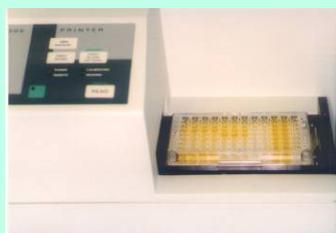
*Mix plate*



*Incubate*



*Wash plate*



*Read plates in a Plate Reader within 30 minutes of the addition of Stop Solution*

step three times. Alternatively, perform these four washes (300  $\mu$ L/well) with a microtiter plate or strip washer. Slap the inverted plate on a paper towel to remove as much liquid as possible.

5. Add **100  $\mu$ L** of **Substrate** to each well.
6. Thoroughly mix the contents of the wells, as in step 2. Cover the wells with new tape or Parafilm and incubate at ambient temperature according to the table below. Use orbital shaker if available.

<b>HIGH SENSITIVITY PROTOCOL</b>	<b>30 minutes</b>
<b>RAPID PROTOCOL</b>	<b>15 minutes</b>

**Caution: Stop Solution is 1.0N Hydrochloric acid. Handle carefully.**

7. Add **100  $\mu$ L** of **Stop Solution** to each well and mix thoroughly. This will turn the well contents yellow.

**NOTE:** Read the plate within 30 minutes of the addition of Stop Solution.

## How to Interpret the Results

### Spectrophotometric Measurement

1. Set the wavelength of the microtiter plate reader to 450 nanometers (nm). (If it has dual wavelength capability, use 600, 630 or 650 nm as the reference wavelength.)
2. Set the plate reader to blank on the Wash/Extraction Buffer Blank wells (this should automatically subtract the mean optical density (OD) of the Blank wells from each control and sample OD). If the reader cannot do this, it must be done manually.

*Note:* The mean OD of the BLANK wells should not exceed 0.2.

### Interpret the Results

A sample is generally considered to be positive if the blank-subtracted OD is greater than or equal to 0.2. For best results, a known negative sample as well as a low-level positive sample should be run as controls in each assay to help judge the results (a conventional and a 0.5% T25 ground corn sample, for example).

**Figure 1A. Example of a typical Qualitative assay setup.**

	1	2	3	4	5	6	7	8	9	10	11	12
A	BL	S6	S14	S22	S30	S38	S46	S54	S62	S70	S78	S86
B	NC	S7	S15	S23	S31	S39	S47	S55	S63	S71	S79	S87
C	PC	S8	S16	S24	S32	S40	S48	S56	S64	S72	S80	S88
D	S1	S9	S17	S25	S33	S41	S49	S57	S65	S73	S81	S89
E	S2	S10	S18	S26	S34	S42	S50	S58	S66	S74	S82	S90
F	S3	S11	S19	S27	S35	S43	S51	S59	S67	S75	S83	BL
G	S4	S12	S20	S28	S36	S44	S52	S60	S68	S76	S84	NC
H	S5	S13	S21	S29	S37	S45	S53	S61	S69	S77	S85	PC



## Precautions and Notes

- Store all Plate Kit components at 4°C to 8°C (39°F to 46°F) when not in use.
- Do not expose Plate Kit components to temperatures greater than 37°C (99°F) or less than 2°C (36°F).
- Allow all reagents to reach ambient temperature (18°C to 27°C or 64°F to 81°F) before use.
- Do not use kit components after the expiration date.
- Do not use reagents or plates from one Kit with reagents or plates from a different Kit.
- Do not expose Substrate to sunlight during pipetting or while incubating in test wells.
- The assay has been optimized to be used with the protocol provided in the kit. Deviation from this protocol may invalidate the results of the test.
- As with all tests, it is recommended that results be confirmed by an alternate method when necessary.
- Observe any applicable regulations when disposing of samples and kit reagents.
- Use extreme caution to prevent sample-to-sample cross-contamination with samples, fluids, or disposables.



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