

### SAMPLE PREPARATION



1. Collect and grind a representative sample such that  $\geq 95\%$  passes through a 20 mesh sieve



Ground too coarse = improper extraction



Ground too fine = extract may require longer settling time

### TEST PROCEDURE

(more detailed instructions in the Product Insert)

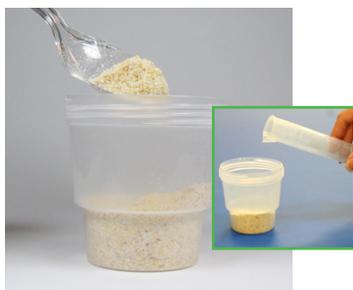
\*use distilled, deionized, or bottled (non-carbonated) water

Note: 2 different pipettes:

- 1 mL adjustable
- 50  $\mu$ L MiniPet

Set out 2 tubes:

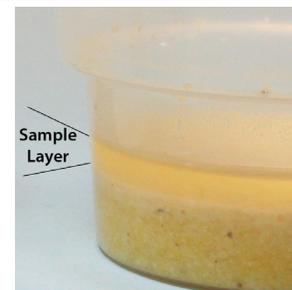
- blue one for mixing,
- clear one for testing



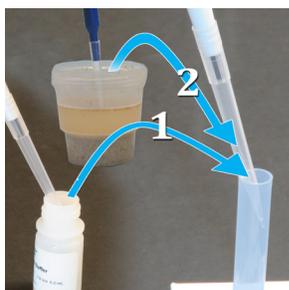
2. Add a 20 or 50 gram sub-sample to container, then add 5 mL of water\* per gram of sample



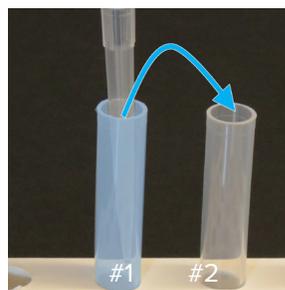
3. Shake 1 minute on mechanical shaker or vigorously by hand for 2 minutes



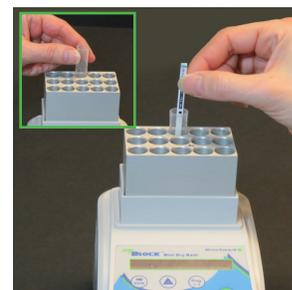
4. Allow to settle into two layers (sample taken from top layer).



5. Using 1 mL pipette and new tip, add 2.5 mL of DB6 Buffer to the first tube (blue dilution tube); then using the MiniPet and new tip, add 50  $\mu$ L extract to the first tube. Mix by pipetting up and down 5-10 times using the 1 mL pipette.



6. Transfer 200  $\mu$ L of the mixed sample from the first tube (blue dilution tube) into the second tube (clear reaction tube)



7. Place tube in Incubator and acclimate for 2 min.\*, then add QuickTox Strip; wait 5 minutes for results. \*Note: acclimation is only required when ambient temperature is unknown or outside of 20-24°C (68-75°F)

### QuickScan TEST RESULTS

(Read single strip(s) alone or along with a QuickComb--more detailed instructions in the QuickScan User Manual)

8. Remove strip from vial immediately after 5 minutes. Cut off and discard bottom pad with arrow tape. (No drying step!)

9. Place in the QuickScan carrier and slide carrier in. Click "Read Test" on Main Menu. Results Screen will appear when scanning is complete.

10. Select "1:A" from Dilution tab pulldown menu on the Results Screen. Base range results are valid between 1.5 and 7 ppm. Enter sample identification data and use buttons to save or print report.