



# DNable<sup>®</sup> Molecular Detection Kit for MON87427 Event Bulk Corn

Part # 12513

## Highlights:

- Molecular Detection of MON 87427 event in Bulk Corn.
- Rapid amplification and detection in a 15 minute assay

## Contents of DNable Kit:

- Reaction Buffer
- MON87427Event Master Mix
- Flat Caps

## Materials Not Provided:

- Pipette capable of delivering 25  $\mu$ L
- Multichannel Pipette capable of delivering 50  $\mu$ L
- Filter Pipette Tips
- Marker
- Extraction Set 10
- AmpliFire DNable Reader\*

\*Available through EnviroLogix

Catalog No. DF-312

## Intended Use

This test kit is intended for rapid qualitative detection of DNA of MON 87427 Event as expressed in bulk corn samples.

## How the Test Works

DNable is an isothermal nucleic acid amplification technology enabling rapid amplification of a specific DNA target. In this test, samples are collected, processed, and the sample extract is added to the reaction buffer. The reaction buffer containing extract is then transferred to the lyophilized Master Mix, containing all the reagents needed to specifically recognize, amplify and detect the MON 87427 Event specific DNA in bulk corn.

The amplified MON 87427 Event specific DNA is detected and the results are displayed and interpreted at 15 minutes using our 8-well DNable Reader.

## Precautions and Notes

DNable is a highly sensitive assay; therefore, the following precautions are recommended to reduce the chance of sample contamination:

- Clean the work stations and pipettes before and after use with 10% bleach solution
- It is recommended to physically separate sample preparation activities from DNable assay activity
- Do not reuse kit disposables
- Use fresh pipette tips for each sample, including replicates from the same sample extract

- Discard used tips in a sealed container containing 10% bleach solution
- Use careful pipetting techniques to avoid cross-contamination between samples; avoid reaching over or pipetting over open tubes
- Wear disposable gloves when handling samples

**Important: Never open reaction tubes after reaction has occurred, as this will release amplified material into the environment and may contaminate subsequent reactions. Care should be taken when disposing of run reaction tubes to avoid possibility of tube leakage. Place completed reaction tubes back in original zippered pouch prior to disposal.**

## Kit Components

- Reaction Buffer:** Provided in green 8-well strip tubes (6)
- MON 87427 Event Master Mix:** Lyophilized reagents provided in clear 8-well strip tubes (6 strips).
- Flat Caps:** used for capping the clear tubes prior to assay start (6 strips)

## Before Testing

- Remove needed DNable Kit reagents from refrigerated storage. Allow reagents to come to room temperature before opening sealed white pouches.
- Pre-heat heat block to 100 °C
- Turn on the 8-well AmpliFire Reader using power button on the right side of the instrument.
- Ensure that all assay reagents, extracted sample, pipettes and Flat Caps are ready for use.



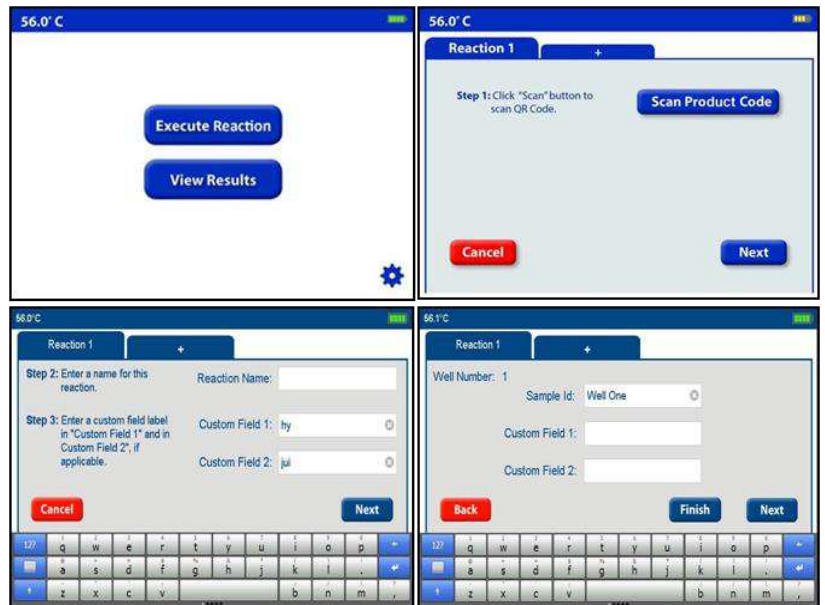
## Sample Preparation

1. Follow Sample Extraction Set 10 product insert for sample preparation, extraction, and thermal denaturation.
2. Remove green Reaction Buffer strip tubes from the kit. Mark the left end tube to note orientation.  
*Important: Tap down or centrifuge green strip tube to ensure that the entire buffer volume is at the bottom of the tubes prior to opening.*
3. Pipette **25 µL of centrifuged sample extract 1** (from Step 1) into well of the reaction buffer 8-well strip tube. Repeat for samples 2-8 in wells 2-8 of the reaction buffer tube, using a fresh pipette tip for each sample.
4. Recap tubes and tap down to ensure all liquid is at the bottom of the tube.

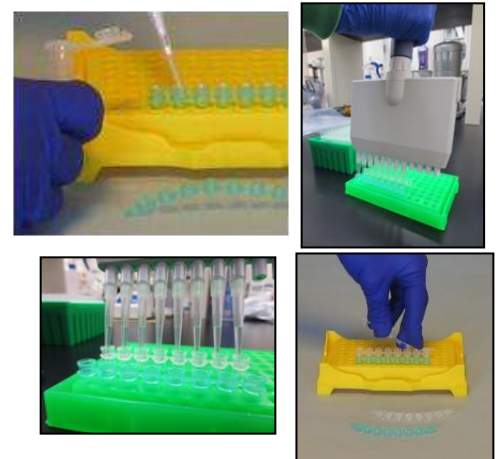
## How to Run the DNable Assay

### DNable assay protocol

1. On the AmpliFire screen, select “**Execute Reaction**” then “**Scan Product Code**”. Use the barcode on the Master Mix foil pouch to scan the MON 87427 Event protocol on the 8-well AmpliFire Reader. “**DF-312**” will display. Select “**Next**”.
2. Under “**Reaction Name**” enter an appropriate reaction description. **This description is placed at the beginning of the file name.** Select “**Next**”
3. To enter specific sample, field or other information for Wells 1 through 8, input information in the Sample ID field and click “**Next**” to advance to each Well. Select “**Finish**” to skip well-specific sample entry.



4. Remove clear Master Mix tubes from the foil pouch and gently tap down to ensure that the white pellet is at the bottom of the tubes.  
*Important: Mark Flat Cap for orientation of the clear Master Mix tubes (writing on clear tubes will interfere with results interpretation or leave marker residue in instrument).*
5. Using a multichannel pipette set to 50 µL, **mix** up and down in the green Reaction Buffer tubes containing sample (1-2-3-4 pipette technique as outlined in the QuickGuide). After mixing, transfer **50 µL** from green tubes to clear Master Mix tubes. Discard clear domed caps from green RB1 Buffer Tubes and Master Mix Tubes. Do **not** mix within the clear tube.



6. **Cap** Master Mix tubes with provided **Flat Caps** strip and mark the left end tube to note orientation.

*Important: Ensure that the tubes are **completely sealed** with Flat Caps as indicated in the picture to the right.*



7. Gently flick down on the resuspended, capped Master Mix to ensure that no bubbles are at the bottom of the tube and that Master Mix is fully resuspended.

8. Inspect tube to ensure that **no air bubbles are present within the sample volume** (a bubble at the top is fine) and that **cap is completely sealed**.

9. When the strip is ready select “**Start**”. Place resuspended, capped clear strip tube in instrument and press “**Ok**”.



10. After 15 minutes, the AmpliFire will produce a short beeping sound and display final results. Results will be interpreted as Not Detected (-), Positive (+) or Invalid (!)\*

**Important:** The full assay time must be complete for result interpretation. (Empty wells will be interpreted as negative.)

\* Invalid denotes that the internal control did not amplify as expected and indicates that either the sample has inhibited the reaction or that something was wrong with the Master Mix or testing procedure.

11. After the assay is complete, carefully **remove run reaction strip tubes from instrument and place in opened foil pouch** (used to store Master Mix), seal and discard in waste container.
12. To export results, return to the home screen, then “View Results”. Insert a USB storage device into instrument (left side) and select each run to export and “Export Selected” and “OK.” The results will be saved in a PDF summary report as well as .csv file format.

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