

Application Note

**Tecan Freedom EVO[®] &
Roche LightCycler[®] 480 II**

DNAble[®]

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ABSTRACT

Genetic trait detection using nucleic-acid based amplification is widely used in agricultural life science applications. DNable® is a rapid, isothermal amplification technology that detects specific nucleic acid sequences with molecular beacons. Similar in mechanism to PCR, DNable has the added flexibility to work with crudely prepared samples, provides a rapid time-to-result, and does not require thermal cycling for amplification. This application note outlines a simple workflow for performing high-throughput DNable testing using the Tecan® Freedom EVO automated liquid handling (ALH) system in conjunction with a Roche LightCycler® 480 II real-time thermocycler. As an industry standard, Artel MVS® was used to calibrate ALH pipetting accuracy prior to DNable testing. Two DNable assays were executed using these platforms; a synthetic singleplex assay that yields quantitative results across a dilution series, and a crude sample (soy leaf) multiplex assay that yields qualitative duplexed results across individual samples. Artel MVS results verified both pipette heads (Span8 LiHa and 96-channel MCA) were dispensing within 5% CV across a 384-well plate. Linear regression ($R^2 > 0.92$) across a 7-log dilution series of target was used to analyze singleplex/quantitative data. Multiplex/qualitative testing across 640 soy leaf samples yielded >95% accuracy with a 95% confidence interval for both assay targets. Total time to result for multiplex testing of 768 DNable reactions (including sample preparation, liquid handling, and thermodetection) was 45 minutes. In conclusion, DNable chemistry offers a rapid method for targeted nucleic-acid amplification in crudely prepared samples that reduces time-to-result by 4-fold compared to standard PCR while maintaining industry-standard accuracy rates (95% sensitivity/specificity with 95% confidence interval).

INTRODUCTION

Genetic trait detection using nucleic-acid based amplification is widely used in agricultural life science applications including field surveillance, adventitious presence testing, and trait confirmation/purity. DNAb^{le}® is a rapid, isothermal amplification technology that uses molecular beacons to specifically detect amplified nucleic acid sequences. The DNAb^{le} system is tolerant of crude sample extracts and therefore no DNA purification is required, removing costly and time-consuming steps from the testing workflow. Since DNAb^{le} amplifies and detects target DNA sequences at a constant temperature, assay results are generated in as little as 5 minutes with excellent analytical sensitivity and specificity. The simple sample preparation combined with the rapid assay run time makes DNAb^{le} effective in low to high throughput testing environments where time to result and testing efficiency are critical for success. For these reasons, DNAb^{le} has great potential to deliver operational efficiencies in seed breeding and seed production workflows.

This application note outlines a simple workflow for performing high-throughput DNAb^{le} testing across two different assay systems using the Tecan Freedom EVO® automated liquid handler and Roche LightCycler® 480 II thermodetection system.

HTP – High throughput; refers to any level of testing > 15,000 reactions per day

MM – Master Mix (resuspended liquid form of the molecular assay)

MVS – Multichannel Verification System (Artel®)

PM – Preventative maintenance (process required to ensure normal operation)

ALH – Automated liquid handler

MATERIALS AND METHODS

1 | Instrumentation & Workflow

To perform automated liquid handling (ALH) of both sample and assay reagents we used a Tecan Freedom EVO ALH equipped with two dispense heads; 1) an 8-channel Spanner LiHa (Span8) used to multi-dispense master mix, and 2) a 96-channel air-dispense pipet head (MCA) used to single-dispense sample extracts. Following industry standards, optimization of liquid class in the Tecan software was necessary to ensure accurate and reproducible pipetting; see Appendix A for specific setting recommendations. To maintain reaction temperature and capture fluorescence data resulting from amplification we used a Roche LightCycler® 480 II. Raw data was collected using LightCycler 480 v1.5.1.62 and analysis was performed in Excel 2016.

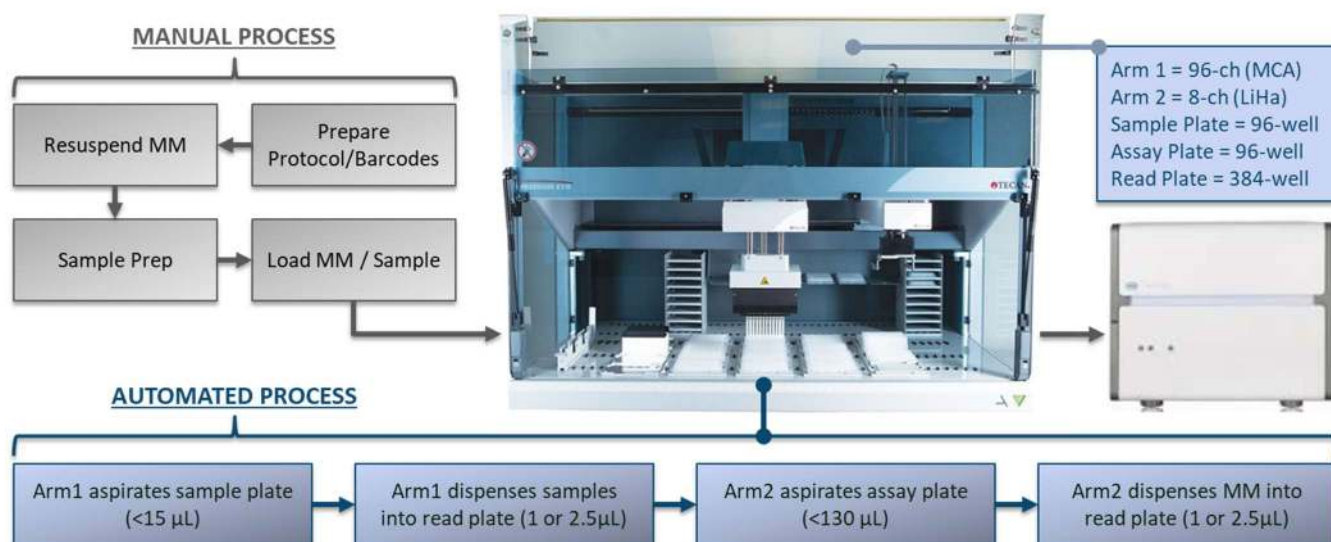


Figure 1. High-throughput workflow for DNAbLe using Tecan Freedom EVO and Roche LightCycler 480 II.

2 | DNAbLe® Assay Reagents

Two DNAbLe assay systems were tested using the Freedom EVO ALH paired with the LightCycler 480 II. The first was a singleplex, quantitative assay using a synthetic oligonucleotide target and the second was a multiplex, qualitative assay using freshly prepared crude soy leaf extract. The singleplex assay produces a standard curve across a seven-log dilution series of synthetic target in less than eight minutes of amplification whereas the multiplex assay produces absence/presence results in crudely prepared samples within fifteen minutes. For both assays, DNAbLe reagents were prepared as lyophilized material requiring only the addition of Reaction Buffer to resuspend active components.

3 | Samples for Testing

For singleplex assays, known concentrations of oligonucleotide were used to generate a seven-log dilution series ranging from 1×10^6 copies/ μL to 1×10^0 copies/ μL . For the multiplex assays, Soy leaf tissue known to be Trait A positive, Trait B positive, or Trait A/B positive were used. Leaf samples (collected as punches) were lyophilized (for shipping only; not required) and processed on-site by the addition of 200 μL of MB15 extraction buffer followed by heating @ 95°C for 5 minutes in a standard heat block. For users lacking the proper format heat block, an alternative heating step has also been validated, which utilizes a simple dry-oven set to 95°C. Crude extracts were then cooled to room temperature before being used for testing.

4 | Artel® Volumetric Evaluation (MVS)

The Artel MVS® system was used to assess the volumetric accuracy of dispensing 2.5 μL of MasterMix or sample using either head (Span8 and MCA) on the Tecan EVO. All testing used Range B solution (Artel) to determine accuracy of dispense across a 384-well plate, resulting in 4 replicates per channel (MCA) or 48 replicates per channel (Span8). All plates were read using the Artel MVS® System running MVS® Data Manager 3.2 Advanced software. Volumetric verification via the Artel MVS system was crucial to ensure proper pipetting accuracy as a function of dispense pattern, liquid class characteristics, and environmental conditions.

RESULTS

5 | Artel® Volumetric Evaluation (MVS)

Dispense Head	Tip Type	# of Replicates	# of Tests	Target Volume (μL)	Mean Volume (μL)	Inaccuracy (%)	CV (%)
Span8	Tecan LiHa Filtered/50 μL	48 per channel	384	2.50	2.46	-1.78 %	4.98 %
MCA	Tecan MCA Filtered/50 μL	4 per channel	384	2.50	2.55	2.04 %	4.14 %

Table 1. Artel MVS summary of volumetric accuracy testing for Span8 and MCA heads on Tecan EVO.

6 | DNable® Verification & Validation

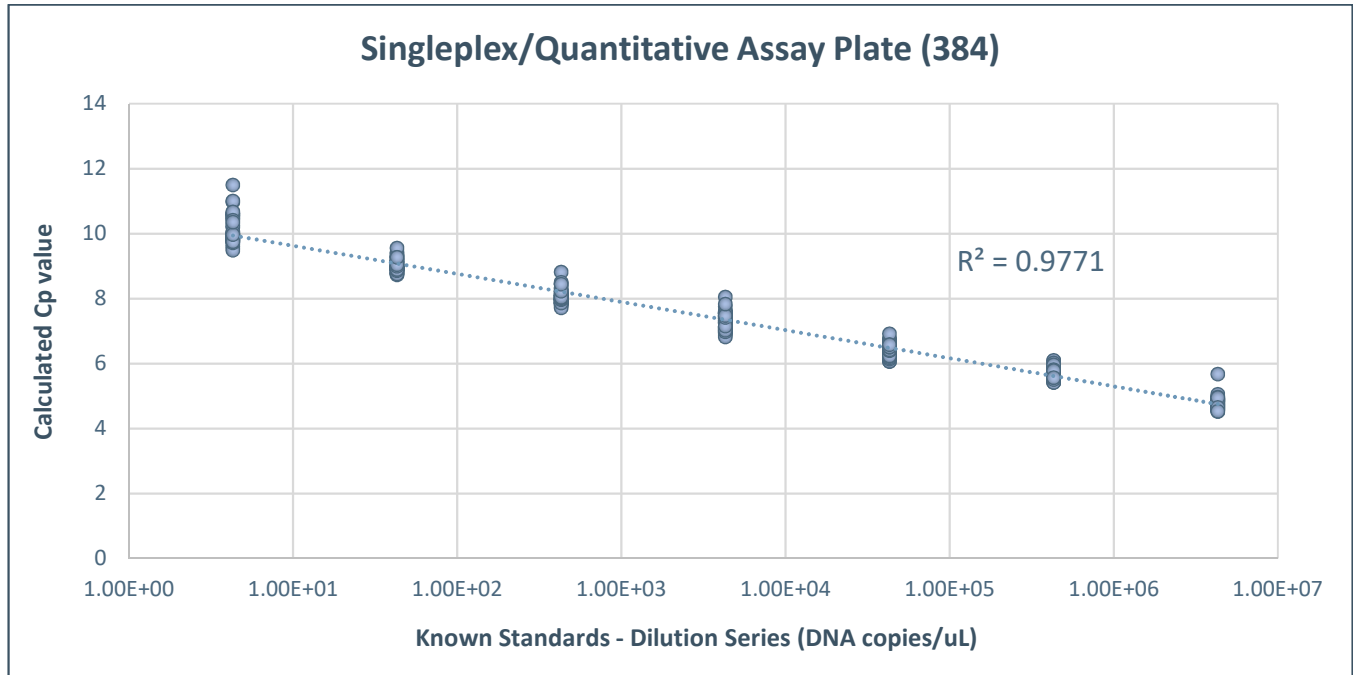


Figure 1. Representative standard curve of Singleplex/Quantitative Assay results for one 384-well plate. Samples were tested as a 7-log dilution series of synthetic target. Across all ten, 384-well test plates, the average R^2 value was 0.924.

Trait/Sample Condition	Biological Replicates	Technical Replicates	Total # of Rxns	ALLELE A (RR2) Accuracy (No CI)	ALLELE A (RR2) Accuracy (95% CI)	ALLELE B (CONV) Accuracy (No CI)	ALLELE B (CONV) Accuracy (95% CI)
NTCs	160	4	640	98.1%	97.0%	100%	99.5%
Conventional (-/-)	160	4	640	99.8%	99.3%	100%	99.5%
Heterozygous (+/-)	160	4	640	100%	99.5%	100%	99.5%
RR2 (+/+)	160	4	640	100%	99.5%	100%	99.5%

Table 2. Multiplex/Qualitative Assay Results for ten 384-well plates. Samples were tested as 16 biological replicates per condition, with four technical replicates, per test plate. Both assays in this multiplex system achieved >96% accuracy with a 95% confidence interval across all ten test plates.

Assay System	Method	Sample Prep	Liquid Handling	Manual Labor	Detection	Total Time
Singleplex DNable	N/A	5 min	12 min	2 min	8 min	27 min
Multiplex DNable	Heat Block	15 min	12 min	2 min	15 min	44 min
Multiplex DNable	Incubator	50 min	12 min	2 min	15 min	79 min
Standard PCR	DNA Purification	60-120 min	12 min	2 min	50 min	124-184 min

Table 3. Process, Throughput, and Cost Comparison between Singleplex, Multiplex, and PCR workflows. Significant time savings in a commercial environment can be realized with DNable due to the rapid sample preparation and reduced thermodetection requirement.

DISCUSSION

In conclusion, DNable chemistry offers a rapid method for targeted nucleic-acid amplification in crudely prepared samples that reduces time-to-result by 4-fold compared to standard PCR while maintaining industry-standard accuracy rates (>95% sensitivity/specificity with 95% confidence interval). This time advantage over standard PCR, along with the cost savings associated with crude sample preparation, leads to significant gains in a commercial testing environment where efficiency is key. Additionally, DNable technology provides flexibility in point-of-detection for field-based applications; users are now able to survey fields in real-time, producing actionable data that can be leveraged within the same day as opposed to sending out samples for third party and/or laboratory-based testing.

REFERENCES

1. J.T. Bradshaw, T. Knaide, A. Rogers, R. Curtis, JALA, 2005, 10 (1), 35-42. "Multichannel Verification System (MVS): A Dual-Dye Ratiometric Photometry System for Performance Verification of Multichannel Liquid Delivery Devices."

APPENDIX

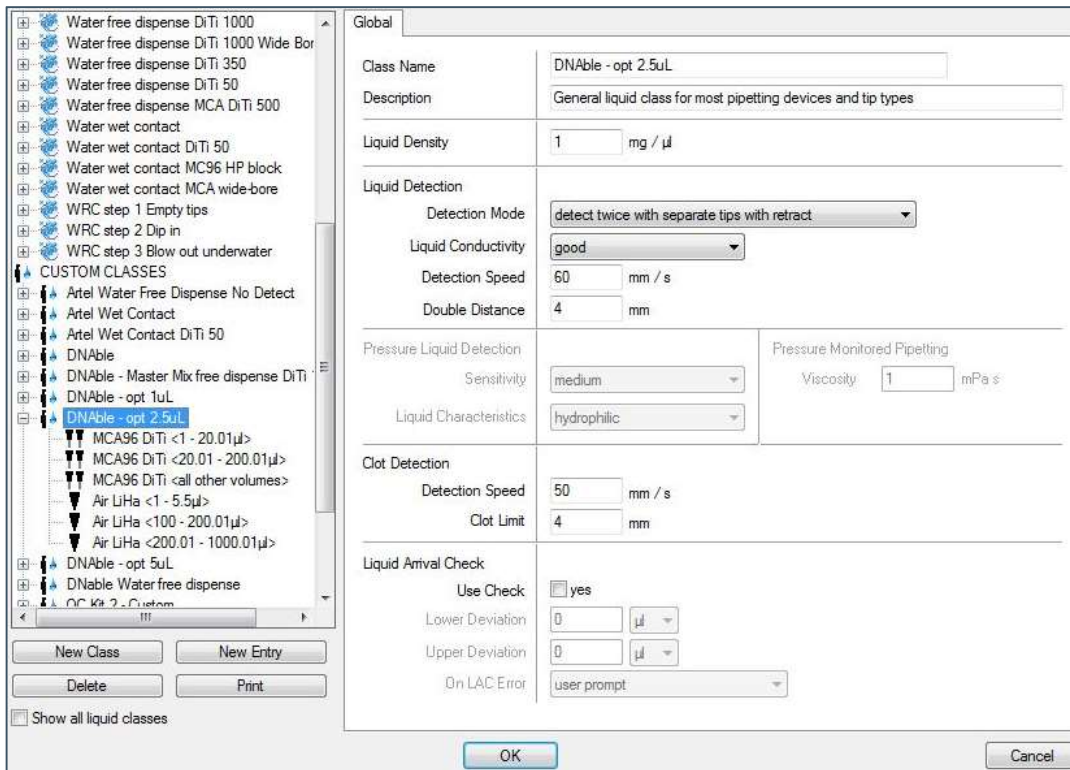


Figure S1. Liquid Class Characteristics - Global Settings

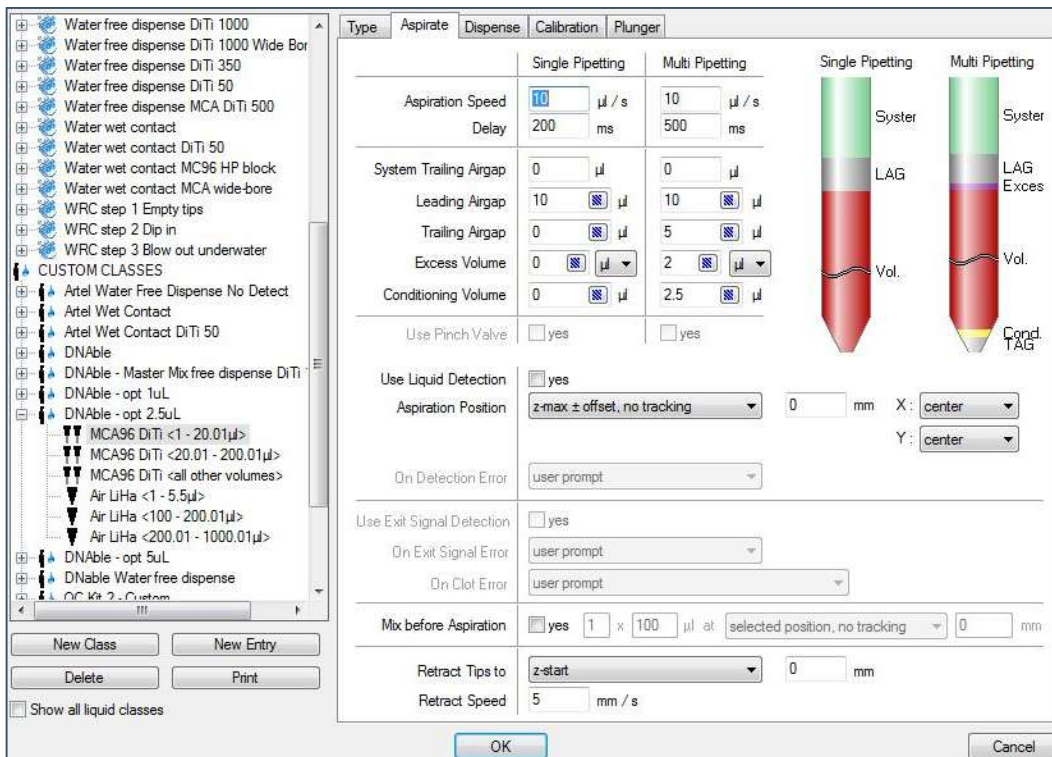


Figure S2. MCA (96-ch) – Aspiration Settings

Dispense Settings

Parameter	Single Pipetting	Multi Pipetting
Dispense Speed	500 µl / s	500 µl / s
Breakoff Speed	µl / s	µl / s
Delay	200 ms	200 ms
Trailing Airgap after each Dispense	<input checked="" type="checkbox"/> yes	<input checked="" type="checkbox"/> yes
Use Pinch Valve	<input type="checkbox"/> yes	<input type="checkbox"/> yes
Use Liquid Detection	<input checked="" type="checkbox"/> yes	
Dispense Position	z-dispense ± offset, no tracking	
Tip Touching	move tips right	
Speed	10 mm / s	
Delay after touching	200 ms	
Mix after Dispense	<input checked="" type="checkbox"/> yes	
Retract Tips to	current position	
Retract Speed	42 mm / s	

MCA (96-ch) – Dispense Settings

Calibration Settings

Offset: 0.12 µl
Factor: 1

Single Pipetting

↑ diluter volume

desired volume

1.0µl 20µl

Multi Pipetting

↑ diluter volume

desired volume

1.0µl 20µl

MCA (96-ch) – Calibration Settings

Mode	Accelerate with (μL/s²)	Decelerate with (μL/s²)	Aspirate (μL/s²)	Dispense (μL/s²)
Single Pipetting	5760	16000	6000	16000
Multi Pipetting	5760	16000	24000	30000

Dispense Speed Curve: 93.4 μ/s Speed vs Time 21ms. Dispense volume: 1.0 μL. Mode: Single Pipetting.

MCA (96-ch) – Plunger Settings

Parameter	Single Pipetting	Multi Pipetting
Aspiration Speed	20 μL/s	20 μL/s
Delay	400 ms	400 ms
System Trailing Airgap	0 μL	0 μL
Leading Airgap	10 μL	10 μL
Trailing Airgap	3 μL	0 μL
Excess Volume	0 μL	5 μL
Conditioning Volume	0 μL	0 μL
Use Pinch Valve	<input type="checkbox"/> yes	<input type="checkbox"/> yes
Use Liquid Detection	<input checked="" type="checkbox"/> yes	
Aspiration Position	z-max ± offset, no tracking	3 mm
On Detection Error	user prompt	
Use Exit Signal Detection	<input type="checkbox"/> yes	
On Exit Signal Error	user prompt	
On Clot Error	user prompt	
Mix before Aspiration	<input checked="" type="checkbox"/> yes	1 x 100 μL at selected position, no tracking
Retract Tips to	z-travel	-5 mm
Retract Speed	20 mm/s	

Diagrams show Single Pipetting (TAG) and Multi Pipetting (LAG, Exces, Vol.) with corresponding aspiration settings.

LiHa (8-ch) – Aspiration Settings

Dispense Settings

Parameter	Single Pipetting	Multi Pipetting
Dispense Speed	800 $\mu\text{l} / \text{s}$	400 $\mu\text{l} / \text{s}$
Breakoff Speed	400 $\mu\text{l} / \text{s}$	400 $\mu\text{l} / \text{s}$
Delay	0 ms	0 ms
Trailing Airgap after each Dispense	<input type="checkbox"/> yes	<input type="checkbox"/> yes
Use Liquid Detection	<input type="checkbox"/> yes	
Dispense Position	z-dispense \pm offset, no tracking	
Offset	5 mm	
X Position	center	
Y Position	center	
Tip Touching	move tips left	
Speed	5 mm / s	
Delay after touching	200 ms	
Mix after Dispense	<input type="checkbox"/> yes	
Mix Volume	1 x 100 μl at selected position, no tracking	
Mix Distance	0 mm	
Retract Tips to	z-dispense	
Retract Speed	50 mm / s	

LiHa (8-ch) – Dispense Settings

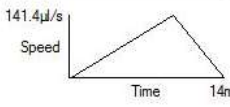
Calibration Settings

Offset: -0.269 μl
Factor: 1.29

Individual settings for dilutors which are physically installed on the instrument:

Tip	Individual Calibration	Offset μl	Factor
1	<input type="checkbox"/>	-0.269	1.29
2	<input type="checkbox"/>	-0.269	1.29
3	<input type="checkbox"/>	-0.269	1.29
4	<input type="checkbox"/>	-0.269	1.29
5	<input type="checkbox"/>	-0.269	1.29
6	<input type="checkbox"/>	-0.269	1.29
7	<input type="checkbox"/>	-0.269	1.29
8	<input type="checkbox"/>	-0.269	1.29

LiHa (8-ch) – Calibration Settings

Type	Aspirate	Dispense	Calibration	Plunger
Single Pipetting	Aspirate	Dispense		
Accelerate with	5000 $\mu\text{/s}^2$	15000 $\mu\text{/s}^2$		
Decelerate with	5000 $\mu\text{/s}^2$	30000 $\mu\text{/s}^2$		
Multi Pipetting	Aspirate	Dispense		
Accelerate with	5000 $\mu\text{/s}^2$	30000 $\mu\text{/s}^2$		
Decelerate with	5000 $\mu\text{/s}^2$	30000 $\mu\text{/s}^2$		
Dispense Speed Curve				
		Dispense volume	1.0 μl	
			Single Pipetting	
Backlash compensation				
<input type="checkbox"/> Sample	2	μl		
<input type="checkbox"/> Airgap	2	μl		

LiHa (8-ch) – Plunger Settings

CONTACT US



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