

TotalTox™ Aflatoxin

Catalog AQ 309 BG

Part #12376

Matrices and Detection Ranges:

(Note: This lot is not enabled for MG3, high-sensitivity peanut)

			Limit of	Highest
		Results reported in	Detection	Approved
Matrix Group ID	Protocol	the range of:	(LOD)*	Level*
	Base Range	0 - 30 ppb	2.7 ppb	30 ppb
AF MG1 - Corn	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Dilution B	0 - >300 ppb	100 ppb	300 ppb
	Base Range	0 - 30 ppb	3.0 ppb	30 ppb
AF MG2 - DDGS	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Dilution B	0 - >300 ppb	100 ppb	300 ppb
	Base Range	0 - 30 ppb	3.0 ppb	30 ppb
AF MG4 - Sorghum	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Dilution B	0 - >300 ppb	100 ppb	300 ppb
AF MG5 – Masa flour	Base Range only	0 - 30 ppb	2.7 ppb	30 ppb
AF MG6 – Corn flour	Base Range only	0 - 30 ppb	2.7 ppb	30 ppb
AEMCZ Drown rise	Base Range	0 - 30 ppb	2.7 ppb	30 ppb
AF MO7 – Brown rice	Dilution A	0 - >100 ppb	30 ppb	100 ppb
AEMC9 Wheet	Base Range	0 - 30 ppb	2.7 ppb	30 ppb
AF MO8 – wheat	Dilution A	0 - >100 ppb	30 ppb	100 ppb
AF MG9 – Corn (High sensitivity)	Base Range only	0 - 10 ppb	1.5 ppb	10 ppb
	Base Range	0 - 30 ppb	2.5 ppb	30 ppb
AF MG10 – Corn Germ	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Dilution B	0 - >300 ppb	100 ppb	300 ppb
AEMG12 Com Gluton Mool	Base Range	0 - 30 ppb	2.5 ppb	30 ppb
AF MO12 – Colli Oluteli Meal	Dilution A	0 - >100 ppb	30 ppb	100 ppb
AE MG13 Corr Gluten Feed	Base Range	0 - 50 ppb	2.7 ppb	50 ppb
AF MOTS – Com Gluten Feed	Dilution A	0 - >200 ppb	50 ppb	200 ppb
AF MG15 Souhean Meal	Base Range	0 - 30 ppb	2.5 ppb	30 ppb
AF MOT5 – Soybean Mean	Dilution A	0 - >100 ppb	30 ppb	100 ppb
AFMG17 Cottonsood (delinted)	Base Range	0 - 30 ppb	2.5 ppb	30 ppb
AF MOT7 – Cottoliseed (definited)	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Base Range	0 - 30 ppb	2.5 ppb	30 ppb
AF MG18 – Barley	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Dilution B	0 - >300 ppb	100 ppb	300 ppb
	Base Range	0 - 30 ppb	2.5 ppb	30 ppb
AF MG21 – Rye	Dilution A	0 - >100 ppb	30 ppb	100 ppb
	Dilution B	0 - >300 ppb	100 ppb	300 ppb
AFMG22 = Oats	Base Range	0 - 30 ppb	2.7 ppb	30 ppb
Al WIG22 - Oats	Dilution A	0 - >100 ppb	30 ppb	100 ppb
AF MG26 – Corn Fermented Protein	Base Range	0-20 ppb	3 ppb	20 ppb
AF MG27 – Peanut Seed	Base Range	0 - 30 ppb	2 ppb	30 ppb
AF MG28 – Peanut Oil	Base Range	0 - 30 ppb	4 ppb	30 ppb

*Do not assume accuracy for results reported below the protocol's LOD or above the protocol's highest approved level.

Important Notes:

- This kit has been certified as a *Performance Tested Method*SM, #012104 by the AOAC Research Institute for use in corn.
- Before testing, the enclosed Multi-Matrix Barcode Card (MMBC) must be scanned just once for each kit lot to upload information to the QuickScan
- QuickScan Software Version 5.9 or later is required





A Summary Guide for testing is provided on Page 15-19. More details for each step in the process are described below and are important for achieving optimal, accurate results.

Matrices

Note: Scanning the Multi-Matrix Barcode Card once per kit lot is required. The QuickScan software will prompt users to select a Matrix Group (MG) before proceeding to the result screen. If you only plan to test matrices within the MG1 group (Corn), scan the side of the MMBC card that only has the MG1 barcode. This allows the software to skip the Matrix Group selection step. Corn, Sorghum, Brown rice, EB17 Buffer Extraction **SET A PROCEDURE: PAGE 6** Wheat, Barley, Oats DDGS, Corn germ, Corn gluten meal, Corn high sensitivity, Rye, 50% Ethanol Extraction **SET B** PROCEDURE: PAGE 7 Soybean Meal, Cottonseed, Corn Fermented Protein Masa flour, Corn flour **SET C PROCEDURE: PAGE 8** EB17 Buffer Special Extraction **SET D** PROCEDURE: PAGE 9 Corn Gluten Feed 84% Acetonitrile Peanut Seed, Peanut Oil Salt Water & 80% Ethanol Mix **SET E PROCEDURE: PAGE 10**

Intended Use

TotalTox Aflatoxin is designed to quickly provide quantitative results for the presence of total aflatoxins. Please refer to Matrix Groups table on page 1 for the Limit of Detection (LOD) and Assay Range(s) for each matrix.

Contents of Kit:	*Available Access	ories:	
• 50 TotalTox Strips packed in a moisture-	Item	Catalog No.	Part #
resistant canister	QuickScan TM System	ACC 351	13065
• 50 EB17 dissolvable pouches (1 pkt per 25g sample [per 10g sample for flour matrices])	5 oz Sample cups/lids	20-0047	10167
 50 reaction tubes 100 pipette tips (1-200 μL) 	<i>Case of 500; for extrac</i> <i>cups with an acetonit</i> <i>covers onto cups with</i>	ting samples up to 30g. Note: 1f usi trile extraction, they may leak; s h Parafilm or similar sealant	ng these eal
DB5 BufferMulti-Matrix Barcode Card - kit lot specific	10 oz Sample cups/lids Case of 100; for extrac	20-0129 ting samples >30g	12383
Hama Not Duaridad.	Graduated cylinder (10	0 mL) ACC 068	11207
items Not Provided:	MiniPet pipette (100 µ	L) ACC 041	11202
 QuickScan System[*] Incubator (base + block)* 	Coffee filters (100)	ACC 083	11434
 Bunn grinder or equivalent 20 mash scream (available through Seadhure or 	Centrifugation Set: Disposables for 50 tests	ACC 010	11214
• 20-mesh screen (available through sectouro of other vendor)	Microcentrifuge	ACC 064 E	11204
Digital scale for weighing samplesExtraction cups with lids* or other suitable	50g Sample Extraction Additional EB17 dissol	Set ACC 099 vable pouches and sample cups (10	12409 0)
vessels for sample extraction	50% Ethanol	ACC E26902-1X	11156
Graduated cylinder*Orbital/rotary shaker	DB5 Buffer Add'l Buffer needed j	KR-266-7 for matrices requiring $> 100 \ \mu L$	11665 per Strip
 Pipette to deliver 100 µL* Pipette to deliver larger volumes 	Dilution Set: Blue dilution tubes and	ACC 103 EB17 dissolvable pouches for 50 te	12500 ests
 Ethanol 50%* (Reagent Alcohol, for some matrices) 	Dilution Tubes: Blue dilution tubes for t	ACC 098 non-EB17 dilution, 50	12236
• Ethanol 80% (Reagent Alcohol for some	1 mL adjustable pipette	e ACC 1303-PRO-1000	11964
matrices)	Pipette tips for 1 mL		
• Table Salt (for some matrices)	pipette (50)	20-0127	12243
• Acetonitrile 84% (for some matrices)	Incubator	ACC BSH301	12458
• Timer			
Scissors			
• Distilled, deionized or bottled water			



How the Test Works

A composite sample is collected, ground, and extracted to solubilize any aflatoxin present. The extract is further diluted into correct buffer before being run on the TotalTox test strip. Each TotalTox Strip has an absorbent pad at each end. The sample extract travels up the test strip and is absorbed into the larger pad at the top of the strip. At the end of the reaction time, the strip is cut at the top of the arrow tape, the bottom pads are discarded, and the strip is inserted into the QuickScan reader to obtain quantitative results.

Matrix specific extractions and analysis protocols are chosen for accuracy and precision. Each matrix is assigned to a Matrix Group (MG). Each MG has a common standard curve, Limit of Detection (LOD), and maximum reported value. When the user selects the MG during testing, the QuickScan System software reads the test strip, retrieves information encoded in the strip's barcode and on the Multi-Matrix Barcode Card (MMBC), and uses the appropriate curve to obtain a result for the matrix being tested.

Precautions – Read First!

SAFETY

- 1. **Disposal of aflatoxin-contaminated materials.** Follow your facility's safety procedures for disposal of samples and extracts potentially containing or known to contain aflatoxin.
- 2. EB17 Dissolvable Pouches contain powder that is flammable and an irritant. See attached Safety Data Sheet.
 - a. If the pouches are damaged, avoid inhaling powder or contact with the skin, eyes, or clothing. Wear personal protective equipment including safety glasses, gloves, mask and lab coat when handling. Keep powder away from heat, sparks and open flame.
 - b. Observe any applicable regulations when disposing of extracted samples and kit reagents.
 - c. Do not treat either the EB17 extracts or the EB17 extraction labware with bleach; the Extraction Pouch powder is incompatible with strong oxidizers.
- 3. Ethanol is flammable and toxic.
 - a. Avoid inhaling vapors or contact with the skin, eyes, or clothing. Wear personal protective equipment including safety glasses, nitrile gloves (not latex), a vapor mask and a lab coat when handling. Keep containers tightly closed and away from heat, sparks and open flame.
 - b. Observe any applicable regulations when disposing of samples and kit reagents.
- 4. Acetonitrile may leak.
 - a. Use caution with extraction cups, assure a tight seal.
 - b. To avoid leaks when using Sample Cups (20-0047), wrap Parafilm[®] or similar product around the outside cup threads in the direction of the threads before capping.

GENERAL

- 1. The intended user should read the entire product instructions, including all safety precautions, before use of this kit. The operator should be capable of using common testing equipment including an appropriate grinder or mill, pipettes, graduated cylinders, etc. Training on use of this product and the QuickScan System is available from EnviroLogix.
- 2. Test strip canisters are desiccated; before opening canisters, ensure they have warmed to room temperature. After removing test strips, reseal the canister immediately. Avoid bending test strips.
- 3. Ensure all samples, extraction reagents (including water), test strips, and Buffer are at room temperature before use.
- 4. As soon as water is added to the sample containing dissolvable EB17 pouches, the sample must be shaken immediately in a hard-walled container to prevent the extraction powder from clumping and not going into solution.
- 5. Test extracts within 5 minutes of diluting with Buffer for optimal performance.

Sample Preparation

- 1. Collect a composite sample according to your own sampling plan or FGIS guidelines. Consult USDA/AMS/FGIS/GIPSA reference documents to help design a plan that fits your needs. Contact Technical Support for more information.
- 2. Grind samples to provide a consistency such that 95% passes through a 20-mesh sieve. Note: No need to further grind masa flour or corn flour. Note: Wheat should be ground such that 70-80% passes through a 20-mesh sieve.
- 3. Mix ground material thoroughly before sub-sampling, to minimize variability.
- 4. Weigh 25g or 50g samples (or 10g flour or corn fermented protein samples) into containers that will allow enough head room for the liquid to move forcefully when shaken vigorously. EB17-extracted matrices require **hard-walled** containers.



Sample Clarification

Depending on the sample matrix, there may be multiple acceptable methods for removing particulate from the extract.

	Centrifugation		Filtration
1.	Fill a microcentrifuge tube with extract.	1.	Add an approved coffee filter (e.g. BUNN Part
2.	Centrifuge for the specified time at 2000 x g (rcf, <i>not rpm</i>).		#BUNBCF100B) to a clean vessel.
3.	Use the top layer of extract for all matrices except flour;	2.	Pour extract into filter. Wait no more than 2 min.
	there may be a white floating layer above that extract that	3.	Pull back the filter to access the filtered extract.
	should not be used for testing.		

Testing in Base Range

Refer to Matrix Group instructions (pages 6-10) or Summary Table (pages 15-19) for base range testing.

Range with Dilution

If, after running and reading the test, the initial result is greater than the upper end of the Base Range, samples can be diluted and retested to extend quantitation (see table on page 1). Combine extract with the appropriate Dilution Reagent to create a diluted extract. Measure carefully and mix well.

Dilution Reagents

Corn/MG1, Sorghum/MG4, Brown rice/MG7, Wheat/MG8, Barley/MG18, Oats/MG22

EB17 Dilution Solution: Dissolve 1 EB17 pouch in 150 mL of water and mix well; Dilution Solution mixture will appear cloudy. Label, date, and document the preparation. Dilution Solution can be stored at ambient temperature for 30 days. Thoroughly mix before use.

DDGS/MG2, Corn Germ /MG10; Corn Gluten Meal/MG12, Rye/MG21

50% Ethanol

Corn Gluten Feed/MG13

84% Acetonitrile

Dilution A: For testing samples at levels greater than 30 ppb (> 30 ppb in Base Range)

- A1. Mix 400 μL Dilution Reagent + 100 μL clarified extract in a blue Dilution Tube or other suitable vessel. Save this diluted extract. Note: For Corn gluten meal, use 300 μL Dilution Reagent; for Corn gluten feed, use 500 μL Dilution Reagent.
- A2. Rerun assay as indicated in Base Range but using diluted extract (see pages 5-7). Example: for corn, pipette 100 μ L DB5 + 100 μ L of the diluted extract into a new reaction tube; place tube in 22°C incubator for 2 min⁺, add a new test strip, and wait 4 minutes for test results.
- A3. In the QuickScan Results Screen, use the dilution tab pull down menu to select Dilution A (1:A). The System will adjust and display the aflatoxin level from diluted samples. Adjusted results are valid in the range of **30-100 ppb**.

Dilution B: For testing samples that read greater than 100 ppb in the Dilution A protocol (after selecting 1:A from the Dilution Tab)

- B1. Mix 200 μL Dilution Reagent + 100 μL diluted extract from Step A1 above, in a blue Dilution Tube or other suitable vessel.
- B2. Rerun assay as in Step A2 above.
- B3. In the QuickScan Results Screen, use the dilution tab pull down menu to select Dilution B (1:B). The System will adjust and display the aflatoxin level from diluted samples. Adjusted results are valid in the range of **100-300 ppb**.

^ The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F).

Use of the QuickScan System

Detailed instructions for use of the QuickScan System are supplied with each unit, and can also be found at <u>www.envirologix.com/support</u>. The lot-specific Multi-Matrix Barcode Card (MMBC) must be scanned into the system prior to testing. In summary, a strip is inserted into the reader and the strips are read by touching or clicking on the "Read Test"



area of the screen. The "Select Matrix Groups" screen will appear if more than one barcode was scanned into the system from the MMBC. Select the group that displays the matrix run. Results are then recorded in an electronic worksheet, allowing each user to report and track data easily.

Kit Storage

This TotalTox Kit should be stored refrigerated. Note the shelf life on the kit box. Prolonged exposure to high temperatures may adversely affect the test results; protect all components from extreme hot or cold temperatures. Do not leave in direct sunlight or in a vehicle. Do not open the desiccated canister until ready to use the strips. Prolonged exposure of the test strips to environmental conditions may adversely affect the test results; protect test results; protect test strips from environmental conditions by allowing canister acclimate to room temperature before opening and closing canister as soon as test strips are removed.

Cross-reactivity

The following mycotoxins have been tested with this kit and no false positive results occurred at the 200 ppm level: DON (deoxynivalenol), Fumonisin B₁, Ochratoxin A, Zearalenone.

Notes

- This product is currently not applicable for use in testing any other crops beyond those specified in this Product Insert.
- This assay is calibrated against reference samples supplied by Trilogy Analytical Laboratory, Washington, MO, and other vendors and associated HPLC data.
- As with all screening tests, it is recommended that results be confirmed by an alternate method when necessary.
- The assay has been optimized for use with the protocols provided in the kit. Deviation from these protocols may invalidate the results of the test. Room temperature components, proper and thorough mixing, timing, and accurate pipetting are essential to accurate results.
- The results generated through the proper use of this diagnostic tool reflect the condition of the working sample directly tested. Extrapolation as to the condition of the originating lot, from which the working sample was derived, should be based on sound sampling procedures and statistical calculations which address random sampling effects, non-random sampling effects and assay system uncertainty. A negative result obtained when properly testing the working sample does not necessarily mean the originating lot is entirely negative for the analyte in question.



Set A: EB17-Extracted Matrices

- Review Sample Preparation on page 3 for grinding consistency and notes. Wheat requires a unique grind quality (also noted below).
- Turn on the incubator and set to 22°C for a minimum of 10 minutes before testing. Ensure that the temperature display has stabilized and indicates "OK" before starting the assay. All reagents should be at room temperature.
- Use distilled, deionized, or flat (non-carbonated) bottled water. Drinkable (potable) tap water may be used, with customer validation of water supply. Contact Technical Support to purchase a control set and protocol that can be used to verify your water supply.
- If testing 50-gram samples, additional EB17 pouches are required (50g Sample Set, Catalog No. ACC-099).

Sample Extraction:

-							
Corn, Sorghum, Brown Rice 25g Add 1 EE Add 75 m		Add 1 EB17 pc Add 75 mL wa	B17 pouch to sample nL water		Wet sample immediately by vigorously		
(95% through 20-mesh)	50g	Add 2 EB17 pc Add 150 mL w	ouches to sam	ple	shaking the sam	shaking for 10 seconds.* If needed, shall the sample against the palm of your oth	
Wheat (70-80% through 20-mesh) Barley, Oats (95% through 20-mesh)	25g only	Add 1 EB17 pouch to sample Add 75 mL water			hand or sample next sha	a har areas aking	d surface to loosen up any dry . <u>Immediately</u> proceed to step.
*Shake: choose mechanical shaker <u>or</u> hand shaking (sorghum: mechanical shaker only) Sha			Shaker Table:mix at highestBy Hand:speed (\geq 300rpm) for 1 minutefor 2 minutes		By Hand: shake vigorously for 2 minutes		
Clarify Extract: choose centrifuge <u>or</u> filter (sorghum: filter only; rice, wheat, barley, oats: centrifuge only)		Centrifuge:30 seconds at 2000 x g (rcf, <u>not rpm</u>)Filter:Pour through coffee filter (ACC (more than 2 min		er: Pour through approved ee filter (ACC 083); wait no e than 2 min			
 Combine Buffer and Extract, then Run Test Strips 1. Add DB5 to the Reaction Tube (discard tip). 2. Add clarified extract to the Reaction Tube. 3. Mix thoroughly with extract pipette tip, discard tip. 				TIPS! Get Co • Ful • Av sha	mplete E ly wet sa oid delay king	Extra mple betw	ction s before the next shaking step yeen water addition and
 Place the Reaction Tube in the 22°C incubator; equilibrate for 2 minutes. Note: tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F). 			 As san 	nple whil	d is n e sha	king	
 Add test strip to tube, arrows down, wait 4 minutes (run time). 				For Be Pip	st Perfor	man nd do	ce wn while mixing

- 6. Immediately cut strips at the top of the arrow tape (discard bottom pads).
- 7. Insert strip into QuickScan Reader.
- 8. When prompted, select Matrix Group for the matrix being tested.
- Avoid Contamination
- Use a new Reaction Tube per test
- Keep DB5 capped, when possible

Read strips promptly after run time

Use new pipette tips for each step

Matrix	LOD (ppb)	First	Second	Third	Shake	Clarify	Reaction Tube	Run						
Corn	2.7	25g	1 x EB17	75mL water	1 .									
					1 m1n	T:14*	100 I DD5	4min						
Sorghum	3.0	50-			snaker*	Filter*	100 µL DB5	(5min:						
Brown Rice	2.7	SUg	2 - ED17	2 - ED17	2 - ED17	2 ED17	2 - ED17	2 - ED17	2 - ED17	150ml water	2min by Co	01 Contrifuco*	$\pm 100 \mu L$	barley,
Wheat	2.7	(corn, sorg.,	2 X EDI /	/ 130mL water	150mL water 211111 by	Centriluge	extract	oats)						
Barley	2.5	fice only)			nanu									
Oats	3.0													

 TABLE A: EB17-Extracted Matrix Summary (Base Range)

*Sorghum: Mechanical shaker only; filter only; Rice, wheat, barley, oats: centrifuge only



- Review Sample Preparation on page 3 for grinding consistency and notes.
- Turn on the incubator and set to 22°C for a minimum of 10 minutes before testing. Ensure that the temperature display has stabilized and indicates "OK" before starting the assay. All reagents should be at room temperature.

Sample Extraction:

Matrix	Ratio	25g Samples	50g Samples
DDGS, Corn germ, Corn gluten meal, Rye, Cottonseed (delinted)	4x 50% ethanol	Add 100 mL to sample	Add 200 mL to sample
Soybean Meal	2x 50% ethanol	Add 50 mL to sample	Add 100 mL to sample
Corn High Sensitivity	1.6x 50% ethanol	Add 40 mL to sample	Add 80 mL to sample
Corn Fermented Protein	4x 50% ethanol	10 g sample: 40 mL 50% et	hanol

Shake: choose mechanical shaker <u>or</u> hand shaking *(rye, mechanical only)*

Shaker Table: mix at highest	By Hand: shake vigorously
speed for 1 minute	for 2 minutes

Clarify Extract: Centrifuge for 1 minute at $2000 \times g$ (rcf, <u>not rpm</u>) or filter through approved coffee filter (ACC-083); wait no more than 2 min (<u>DDGS only</u>).

Combine Buffer and Extract, then Run Test Strips

- 1. Add DB5 to the Reaction Tube (discard tip).
- 2. Add clarified extract to the Reaction Tube.
- 3. Mix thoroughly with extract pipette tip, discard tip.
- 4. Place the Reaction Tube in the 22°C incubator; equilibrate for 2 minutes. Note: tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 24°C (68 75°F).
- 5. Add test strip to tube, arrows down, wait 5 minutes (run time).
- 6. Immediately cut strips at the top of the arrow tape (discard bottom pads).
- 7. Insert strip into QuickScan Reader.
- 8. When prompted, select Matrix Group for the matrix being tested.

TIPS!

Get Complete Extraction

- Fully wet samples before shaking
- Assure liquid is moving forcefully though the sample while shaking

For Best Performance

- Pipette up and down while mixing
- Read strips promptly after run time

Avoid Contamination

- Use a new Reaction Tube per test
- Keep DB5 capped when possible
- Use new pipette tips for each step

TABLE B: 50% Ethanol-Extracted Matrix Summary (base range)

Matrix	LOD (ppb)	Extractant	Shake	Clarify	Add to Reaction Tube	Run Time
DDGS (MG2)	3.0					
Corn Germ (MG10), Rye (MG21)	2.5	4x 50%				
Corn Gluten Meal (MG12)	3.5	ethanol		Filter or		
Cottonseed (delinted) (MG17)	2.5		1 min	Centrifuge		
Soybean Meal (MG15)	2.5	2x 50% ethanol	shaker*	1 min at 2000 x g	100 μL DB5 +	5 min
Corn High Sensitivity (MG9)	1.5	1.6x 50% ethanol	2 min by hand			
Corn Fermented Protein (MG26)	3.0	40 mL 50% ethanol		Centrifuge 1 min at 2000 x g		

*Rye: Mechanical shaker only



Set C: Masa Flour, Corn Flour

- Review Sample Preparation on page 3 for grinding consistency and notes.
- Turn on the incubator and set to 22°C for a minimum of 10 minutes before testing. Ensure that the temperature display has stabilized and indicates "OK" before starting the assay. All reagents should be at room temperature.
- Use distilled, deionized, or flat (non-carbonated) bottled water. Drinkable (potable) tap water may be used, with customer validation of water supply. Contact Technical Support to purchase a control set and protocol that can be used to verify your water supply.

Sample Extraction:

Masa flour,	the palm of your other hand or a hard surface to loosen
Corn flour 10g Add 1 EB17 pou	up any dry sample areas. <u>Immediately</u> proceed to next
Add 60 mL wat	shaking step.

Shake: choose mechanical shaker or hand shaking

Clarify Extract: Clarify <u>immediately</u> after shaking; choose centrifuge <u>or</u> filter

Combine Buffer and Extract, then Run Test Strips

- 1. Add DB5 to the Reaction Tube (discard tip).
- 2. Add clarified extract to the Reaction Tube.
- 3. Mix thoroughly with extract pipette tip, discard tip.
- 4. Place the Reaction Tube in the 22°C incubator; equilibrate for 2 minutes. Note: tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 24°C (68 75°F).
- 5. Add test strip to tube, arrows down, wait 4 minutes (run time).
- 6. Immediately cut strips at the top of the arrow tape (discard bottom pads)
- 7. Insert strip into QuickScan Reader.
- 8. When prompted, select Matrix Group for the matrix being tested.

<i>Shaker Table:</i> mix at highest speed (≥ 300rpm) for 1 minute	<i>By Hand:</i> shake vigorously for 2 minutes		
<i>Centrifuge:</i> 1 minute at 2000 x g (rcf, <u>not rpm</u>); poke through white floating layer (if present) to access extract	<i>Filter:</i> Pour through approved coffee filter (ACC 083); wait no more than 2 min		

TIPS!

Get Complete Extraction

- Fully wet samples before the next shaking step
- Avoid delay between water addition and shaking
- Assure liquid is moving forcefully though the sample while shaking

For Best Performance

- Pipette up and down while mixing
- Read strips promptly after run time

Avoid Contamination

- Use a new Reaction Tube per test
- Keep DB5 capped, when possible
- Use new pipette tips for each step

TABLE C: Masa/Corn Flour Matrix Summary (base range)

Matrix	LOD (ppb)	First	Second	Third	Shake	Clarify	Reaction Tube	Run
Masa, Corn Flour	2.7	10g	1 x EB17	60 mL water	1 min – shaker <u>or</u> 2 min – by hand	Filter <u>or</u> Centri- fuge	100 μL DB5 + 200 μL extract	4 min



Set D: Corn Gluten Feed

- Review Sample Preparation on page 3 for grinding consistency and notes.
- Turn on the incubator and set to 22°C for a minimum of 10 minutes before testing. Ensure that the temperature display has stabilized and indicates "OK" before starting the assay. All reagents should be at room temperature.
- Use distilled, deionized, or flat (non-carbonated) bottled water. Drinkable (potable) tap water may be used, with customer validation of water supply. Contact Technical Support to purchase a control set and protocol that can be used to verify your water supply.

Sample Extraction

Corn Gluten Feed 25g Add 35 mL 84% Acetonitrile	Tightly close extraction container. Wet sample immediately, by vigorously shaking for 10 seconds by hand. If needed, shake the sample against the palm of your other hand or a hard surface to loosen up any dry sample areas. Immediately proceed to next shaking step.
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Shake: choose mechanical shaker

Shaker 1	Table: mix	at highest	speed (\geq	300rpm) for 2 minutes
2		and might be		e o o i p i i i	101 - 1111100000

(rcf, not rpm);

Clarify Extract: Clarify immediately after shaking; choose to settle or centrifuge

Combine Buffer and Extract, then Run Test Strips

- 1. Add DB5 to the Reaction Tube (discard tip).
- 2. Add clarified extract to the Reaction Tube.
- 3. Mix thoroughly with extract pipette tip, discard tip
- 4. Place the Reaction Tube in the 22°C incubator; equilibrate for 2 minutes. Note: tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 24°C (68 75°F).
- 5. Add test strip to tube, arrows down, wait 5 minutes (run time).
- 6. Immediately cut strips at the top of the arrow tape (discard bottom pads).
- 7. Insert strip into QuickScan Reader.
- 8. When prompted, select Matrix Group for the matrix being tested.

TIPS!

Settle: 30 seconds

Get Complete Extraction

 Make sure to close extraction vessel tightly to prevent solvent leak

Centrifuge: 30 seconds at 2000 x g

- Fully wet samples before the next shaking step
- Assure sludge is moving forcefully in extraction vessel while shaking

For Best Performance

- Pipette up and down while mixing
- Read strips promptly after run time

Avoid Contamination

- Use a new Reaction Tube per test
- Keep DB5 capped, when possible
- Use new pipette tips for each step

Matrix	LOD (ppb)	Sample size	Extractant	Shake	Clarify	Reaction Tube	Run
Corn Gluten Feed	2.7	25g	35 mL 84% ACN	2 min – shaker	Settle or centrifuge	175 μL DB5 + 25 μL extract	5 min

TABLE D: Corn Gluten Feed



SET E: Salt Water & Ethanol Mix-extracted Matrices

- Review Sample Preparation on page 3 for grinding consistency and notes.
- Turn on the incubator and set to 22°C for a minimum of 10 minutes before testing. Ensure that the temperature display has stabilized and indicates "OK" before starting the assay. All reagents should be at room temperature.

Sample Extraction:

Matrix	Salt Water Ratio	25g Samples	50g Samples	Custom Samples
Peanut Seed (MG27) Peanut Oil (MG28)	29.4g table salt per 100 mL bottled water	 Add 20 mL salt water to sample Mix well, stir slowly Add 50 mL 80% Ethanol 	 Add 40 mL salt water to sample Mix well, stir slowly Add 100 mL 80% Ethanol 	 Add 0.8 mL/g sample salt water to sample Mix well, stir slowly Add 2 mL/g sample 80% Ethanol

Shake: choose mechanical shaker or hand shaking

Shaker Table: mix at highest speed for 1 minute *By Hand:* shake vigorously for 2 minutes

Clarify Extract: Centrifuge for 1 minute at 2000 x g (rcf, <u>not rpm</u>), pour supernatant into separate vessel for storage.

Combine Buffer and Extract, then Run Test Strips

- 1. Add 300 μ L **DB5** to the reaction vial (discard tip).
- 2. Add 100 µL clarified extract to the reaction vial.
- 3. Mix thoroughly with extract pipette tip, discard tip.
- 4. Add test strip to vial, arrows down.
- 5. Wait 4 minutes (run time).
- 6. Immediately cut strips at the top of the arrow tape (discard bottom pads).
- 7. Insert strip, barcode down, into QuickScan reader.
- 8. When prompted, select Matrix Group 27 for Peanut Seed and Matrix Group 28 for Peanut Oil.

TIPS!

Get Complete Extraction

- Fully wet samples before shaking
- Assure liquid is moving forcefully though the sample while shaking

For Best Performance

- Pipette up and down while mixing
- Do not reuse diluted samples
- Read strips promptly after run time

Avoid Contamination

- Use a new Reaction Tube per test
- Keep DB5 capped when possible
- Use new pipette tips for each step

TABLE E: Salt Water & 80% Ethanol-Extracted Matrix Summary (Base Range)

Matrix	LOD (ppb)	Extractant	Shake	Clarify	Add to Reaction Tube	Run Time
Peanut Seed (MG27)	2.0	0.8 mL/g sample salt water to sample	1 min – shaker	Filter or Centrifuge 1 min at	300 μL DB5 +	4
Peanut Oil (MG28)	4.0	Mix well, stir slowly 2 mL/g sample 80% Ethanol	2 min by hand	2000 x g, pour supernatant into separate vessel	100 µL extract	4 min





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•		Revision nr.2 Dated 04/21/2019
• ENVIROLO	GIX	Page n. 1 / 6
-	Safatu data ak	eet
SECTION 1. Identification of t	he substance/mixture and of	eet f the company/undertaking
1 Product identifier		
Trade name: Part number:	Extraction Buffer EB17(11198, 12382)	
.2 Relevant identified uses of the sub-	stance	
•	specified in product literate	ture.
3 Details of the supplier of the safety sheet Manufacturer/Supplier	data	rida Industrial Diana
Information department:	Portland ME 04103, USA Technical Service	iside industrial rawy.
4 Emergency telephone number:	(207) 797-0300	
1 Classification of the substance or n	nixture	
lassification according to OSHA		
272/2008 (CLP):	Flammable Solid category 2	H228 Flammable solid
	Acute Toxicity Inhalation 4	H302 Hamilu ii swaliowed H322 Hamilu ii inhaled H315 Group chini tation
	Skin Irritation category 2 Serious eye damage category 1	H315 Causes skin irritation H318 Causes serious eye damage
	Single Exposure category 3	H335 May cause respiratory irritation
	Aquatic Toxicity-Chronic category 3	H412 Harmful to the environment with long lasting
2 Label elements Labeling according to OSHA 29CFR 1910.1200 and Regulation (EC) 1272/2008		effects
Hazard pictograms :		
Signal word:	Danger	
Hazard statements:	H228 Flammable solid	
	H302 + H322 Harmful if swallov H315 Causes skin irritati	wed or inhaled
	H318 Causes serious eye H335 May cause respirat	e damage. tory irritation.
Description of the sector	H412 Harmful to aquatic	: life with long lasting effects.
Precautionary statements:	P264 Wash hands thorou P273 Avoid release to the	ughly after handling. he environment.
	P280 Wear protective gl P301 + P312 IF SWALLOWED	loves/ eye protection.): Call a POISON CENTER/doctor/physician if you
	feel unwell. P304 + P340 IF INHALED: Ren P305 + P351 + P320 If in France P	move to fresh air and keep comfortable for breathing.
	P303 + P331 + P338 If in Eyes: R remove cont P403 + P233 Store in a well vent	tact lenses if present and easy to do. Continue rinsing. tilated place. Keep container tightly closed
	store in a treat year	, upmy cover
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Softer hazards: SDS: EB17 SDS: EB17 SDS: EB17 SDS: EB17 SDS: ED17	No additional hazards listed OGIX are measures. The measures. The personal protective equip vapors, mile or gas, Assum al Evacuate personnel to a safe a Prevent further leakage or upi draim. Discharge into the our Sweep up and shovel. Pervent gintion sources. Call for assume the prevent further leakage or upi draim. Discharge into the our Sweep up and shovel. Pervent gintion sources. Call for assume the prevent further leakage or upi draim. Discharge into the our Sweep up and shovel. Pervent gintion sources. Call for assume the prevent further leakage or upi the prevent prevent leakage or upi the prevent further leakage or upi the prevent	Revision nr.2 Dated 04/21/2019 Page n. 3 / 6 ment Avoid dust formation Avoid breathing lequate ventilation. Remove all sources of ignition tree. Avoid breathing dast. llage if safe to do so. Do not let product enter irronment must be avoided. tentry into sewers, dike if needed. Eliminate all stance on disposal. vater on the contaminated surface and allow to system. tion 7. For information on PPE refer to Section 8. 13. p away from sources of ignition. Prevent electrostatic not breathe dust. Wear suitable protective clothing. In on, wear suitable repriratory equipment. If ingestic hately and show the container or the label. Avoid bles such as oxidizing agents. ed. Keep container in a cool, well-ventilated area. in Section 1.2 there are no other specific uses verify total substick in graph separable fraction. (OSHA Observe limits for particulate not otherwise regulated. verify ELD tong m ² inhuable particulate. 3 (OSHA Observe limits for particulate (ACCHIT LV) EEH04/2005 Inhalable dust: 10mg/m ² , Respirable dust: 4mg/m ²
Softer hazards: SDS: EB17 SDS: EB17 SDS: EB17 SDS: EB17 SDS: EB17 SDS: EB17 SDS: ED17	No additional hazards listed Control of the second protective equipation of the second protection o	Revision nr.2 Dated 04/21/2019 Page n. 3 / 6 ment Avoid dust formation Avoid breathing lequate ventilation. Remove all sources of ignition. trea. Avoid breathing dust. Ilage if safe to do so. Do not let product enter tronment must be avoided. Lenty of the contaminated surface and allow to system: to a contaminated surface and allow to system. ion 7. For information on PPE refer to Section 8. 13. p away from sources of ignition. Prevent electrostatic not breathe dust. Wear suitable protective clothing. In ion, wear suitable reparatory equipment. If ingestic, lately and show the container or the label. Avoid bles such as oxidizing agents. ed. Keep container in a cool, well-ventilated area. in Section 1.2 there are no other specific uses versimated. Joing of particulate not otherwise regulated. versimate Limits for particulate not otherwise regulated. versimate Limits (ACGIH TLV) EH40/2005 Inhalable dust: (ACGIH TLV) EH40/2005 Inhalable dust: (ACGIH TLV) and at 4mg/m ² gthis material should be equipped with an eyewash and local exbaust or general duttion ventilation. protein abould be determined according to local pressentements.
Softer hazards: SDS: EB17 SDS: EB17 SDS: EB17 SECTION 6. Accidental release I personal precautions, protective equipment and emergency procedu SECTION 6. Accidental release Soften and emergency procedu Environmental precautions: A Reference to other sections: SECTION 7. Handling and stoc I Precautions for safe storage, includ any incompatibilities: Specific end use(s): SECTION 8. Exposure control Lexposure controls Additional information about design technical systems: Components with limit values that re- monitoring at the workplace: xposure controls - Engineering Cont ersonal protective equipment Breathing equipment	No additional hazards listed Constraints POGIX Re measures. Inter: Use personal protective equipal vapors, mist or gas. Assure ad Execute personal to a safe a Prevent further leakage or gat dmins. Discharge into the env Sweep up and shevel. Pervent gintion sources. Call for assure For safe handling refer to Sector For disposal, re	Revision nr 2 Dated 04/21/2019 Page n. 3 / 6 ment Avoid dust formation. Avoid breathing lequate venilation. Remove all sources of ignition. trace avoid breathing dust. Ilage if safe to do so. Do not let product enter tromment must be avoided. tenty: into servers, dike if needed. Eliminate all stance on disposal. water on the contaminated surface and allow to system. tion 7. For information on PPE refer to Section 8. 13. p away from sources of ignition. Prevent electrostatic not breathe dust. Wear suitable protective clothing. In ion, wear suitable protective clothing. In ion, wear suitable constance or the label. Avoid bles such as ossidizing agents. ed. Keep container in a cool, well-ventilated area. in Section 1.2 there are no other specific uses via black Distribution of DFIA Cheerve limits for particulate not otherwise registratic. (OSILA Observe limits for particulate not otherwise registration (OSILA PEL) 10 mg/m ² inhable particulate, 3 mg/m ² registrable dust: 10 mg/m ² , Respirable dust.4mg/m ² g this material should be equipped with an eyewash and local exhauts or general duiton over unitainon. protocols. An approved disposable an conding to local alysis protocols. An approved disposable an conding to local alysis protocols. An approved disposable an conding to local alysis protocols. An approved disposable an entrying.
A) Other hazards: SDS: EB17 Construct of the section of	No additional hazards listed Control of the second protective equip vapors, mist or gas. Assure ad Execute personnel protective equip vapors, mist or gas. Assure ad Execute personnel to a set a Prevent further leakage or gai dmins. Discharge into the env: Sweep up and shovel. Pervent ignition sources. Call for assure Finish cleaning by spreading ver- encate the thrule leakage or gai exactate through the samitary. For safe handling refer to Section introl. Reep away from heat. Keep to disposal, refer to Section rage. Keep away from heat. Keep buildup. Do not inegen buildup. Do not inegen. Reep away from incompati Keep container tightly closs Desides the uses described in Is/personal protection. Is/personal protection. Is/of None required equire For Section sports of the source of the Sodium Lung Sulfate temps of the source of the Sodium Lung Sulfate respiratory conditions using risk am age respirators and com government standards set	Revision nr 2 Dated 04/21/2019 Page n. 3 / 6 ment. Avoid dust formation. Avoid breathing lequate venilation. Remove all sources of ignition. Irradiate venilation. Remove all sources of ignition. Irradiate venilation and the sources of ignition. Irradiate venilation on PPE refer to Section 8. 13. 1. p away from sources of ignition. Prevent electrostatic not breathe dust. Wear suitable protective clothing. In ion, wear suitable reportective clothing. In ion, wear suitable dust. Sources of in Section 1.2 there are no other specific uses 1 DSBA Observe limits for particulate not otherwise regulated. Is migin 3 total dust, 5 migin ¹¹ respirable fraction (OSIFA PEL) 10 mg ¹² maintable particulate, 3 mg ¹² regulated dust. (ACCIII TLV) I Himmetrial should be equipped with an eyewash and local exhaust or general dilution ventilation. protection should be determined according to local alysis protocols. An approved disposable arc-purifying van an NOSI (US) or CEN (EU).
3 Other hazards: SDS: EB17 SECTION 6. Accidental releas SECTION 6. Accidental releas 1. Personal precautions, protective equipment and emergency procedu 2. Environmental precautions: 3. Methods and material for containment and clean up: 4. Reference to other sections: SECTION 7. Handling and sto 1. Precautions for safe storage, includ any incompatibilities: 3. Specific end use(s): SECTION 8. Exposure control 1. Exposure controls Additional information about design Section 8. Exposure control 1. Sepostre controls Components with limit values that n monitoring at the workplace: xposure controls - Engineering Cont ersonal protective equipment Breathing equipment.	No additional hazards listed Control of the personal protective equip vapors, mist or gas. Assure ad Evenuate personnel to a suft a Prevent further leakage or guid dmins. Discharge into the entrol Sweep up and shovel. Prevent jupiton sources. Call for assure Front and the leakage or guid dmins. Discharge into the entrol Sweep up and shovel. Prevent jupiton sources. Call for assure reactate through the samtary. For safe handling refer to Section 1 rage. Keep away from heat. Keep buildup. Do not ingest. Do case of insufficient ventilation scentaectual skine and eyes. Besides the uses described in Issjoersconal protection. Its of None required equire For Section protection. Suffate eventilation of the state source of the source. Solution control of the state solution of Solution compared of the state Solution control of Solution control safety shower. Provide I Appropriate respiratory of the state respiratory and conducter separator and com government standards as and com government standards as and com government standards as and com government standards as and com govern and the spirator and com government standards as a state shower. Provide I Handle with gloves. Cher of the state spirator and com government standards as a state shower. Provide I Handle with gloves. Cher of the state spirator and com government standards as a state shower.	Revision nr.2 Dated 04/21/2019 Page n. 3 / 6 ment. Avoid dust formation. Avoid breathing lequate ventilation. Remove all sources of ignition. urea. Avoid breathing dust. llage if safe to do so Do not let product enter irronment must be avoided. tentry into servers, dike if needed. Eliminate all stance on disposal. water on the contaminated surface and allow to system. tion 7. For information on PPE refer to Section 8. 13. p away from sources of ignition. Prevent electrostatic not breathe dust. Wear suitable protective clothing. In ion, wear suitable reportective clothing. In in Section 1.2 there are no other specific uses the Exposure Limits to SIRA Observe limits for particulate not otherwise regulated. 15 mg/m3 total dust, 5 mg/m1 "respirable fraction (OSIFA PEL) 10 mg/m1 "inhalable particulate, 3 mg/m1 "regurable particulate. (ACCIII TLV) I stafu 2005 Inhalable dust. 10mg/m2, Respirable dust amg/m1 g this material should be determined according to local alysis protocols. An approved disposable air-purifying volume Newlow of under appropriate uota. NNOSH (US) or CEN (EU). via brotosil brief intervia to several duition ventilation.
3 Other hazards: SDS: EB17 SDS: EB17 SECTION 6. Accidental releas 1 Personal precautions, protective equipment and emergency procedur 2 Environmental precautions: 3 Methods and material for containment and clean up: 4 Reference to other sections: SECTION 7. Handling and sto 1 Precautions for safe storage, includ any incompatibilities: 3 Specific end use(s): SECTION 8. Exposure control 1 Exposure controls Additional information about design technical systems: Spoore limits Components with limit values that r monitoring at the workplace: xposure controls - Engineering Cont resonal protective equipment Breathing equipment.	No additional hazards listed CGIX at measures, at measures, res: Use personal protective equip vapors, mist or gas. Assure of Evacuate personnel to a safe a Prevent further leakage or spil dram. Discharge into the envi gintion sources. Call for assure evacuate through the santary. For safe handling refer to Section 1 rage. Keep away from heat. Keep buildup. Do not ingest. Do case of mistificent venthion keep container tightly close Do not ingest. Do case of mistificent venthion Keep away from incompating Response of the same of the same secture of None required equire rols: Facilities using or storin safety shower. Provide 1 Solution for the safety of the safety approximate registratory approximate registratory approximate registratory approximate registratory approximate registratory approximate registratory approximate registratory approximate registratory approximate registratory approximate registratory and compare standards su	Revision nr.2 Dated 04/21/2019 Page n. 3 / 6 ment. Avoid dust formation. Avoid breathing lequate ventilation. Remove all sources of ignition. tree. Avoid breating dust. llage if safe to do so. Do not let product enter womment must be avoided. tatyr into server, dise if needed. Eliminate all stance on disposal. water on the contaminated surface and allow to system. iton 7. For information on PPE refer to Section 8. 13. p away from sources of ignition. Prevent electrostatic nots breath dust. Ware suitable protective coloning: inou, wear suitable respiratory equipment. If ingested, lately and show the container or the label. Avoid bles such as oxidizing agents. d Exposure Limits for particulate not otherwise regulated. (A Keep container in a cool, well-ventilated area. in Section 1.2 there are no other specific uses in given bar of dust 3 mg/m 3 mg/m 3 mg/m 4 mspirmble fraction (a Sing/m 3 Di) al dust 3 mg/m 3 mg/m 4 mspirmble fraction (a Sing/m 3 Di) al dust 4 mg/m 4 mspirmble fraction (a Sing/m 3 Di) al dust 4 mg/m 4 mspirmble fraction (a Sing/m 3 Di) al dust 4 mg/m 4 mspirmble fraction (b Sing/m 3 Di) al dust 4 mg/m 4 mspirmble particulate, 3 imput 4 mspirmble particulate (ACGHT 11V) dust. 4 mg/m 4 upsteriorities. As approved disposable ar-partyling protection should be determined according to local allysis protociols. At approved disposable ar-partyling promets itseled and approved under appropriate uch as NIOSH (US) or CEN (EU).
23. Other hazards: SDS: EB17 SDS: EB17 SECTION 6. Accidental releases 51. Personal precautions, protective equipment and emergency procedur 32. Environmental precautions: 33. Methods and material for containment and clean up: 34. Reference to other sections: SECTION 7. Handling and stoo 31. Precautions for safe storage, includ any incompatibilities: 33. Specific end use(s): SECTION 8. Exposure control 34. Additional information about design technical systems: 35. Spoure limits: Components with limit values that m monitoring at the workplace: Stopoure controls – Engineering Cont Versonal protective equipment Breathing equipment.	No additional hazards listed OGIX ret measures. ret: Use personal protective equip vapor, mist or gas. Assure al Evacuate personnel to a safe a prevent further leakage or upi drains. Discharge into the eav Sweep up and shovel. Pervent ignition sources. Call for assure For safe handling refer to Sectors For discussed and the sector of	Revision nr.2 Dated 04/21/2019 Page n, 3 / 6 ment. Avoid dust formation. Avoid breathing lequate ventilation. Remove all sources of ignition. tree. Avoid breathing dust. llage if safe to do so. Do not let product enter twomment imust be avoided. tentry into severs, dike if needed. Eliminate all stance on disposed stance on disposed. tentry into severs, dike if needed. Eliminate all stance on disposed. tentry into severs, dike if needed. Eliminate all stance on disposed. total > 7 or information on PPE refer to Section 8. 13. p away from sources of ignition. Prevent electrostatic not breathe dust. Ware unstalle preventive clothing. In ion, ware suitable respiratory equipment. If ingested, lately and show the container or the label. Avoid bles such as oxidizing agents. dt Keep container in a cool, well-ventilated area. in Section 1.2 there are no other specific uses vert distance and provol induse particulate not otherwise regulated. ust simplif respiratole particulate (ACGIITLV) EFH0 2005 Inhabile dust. IOmgraf. Respirable dust 4 magina? usin material should be equipped with an eyewash and local exhaust or general dilution ventilation. protect is subckip to engineering controls. Always ponents itseld and approved inder paroprinte un an MOSH (US) or CEN (EU). we must be inspirated particulate after usin thous and good laboratory practices. Wash and dry

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SECTION 3. Composition/information	tion on ingred	lients.			
3.1 Substances: Information not releva	nt				
32 Mixtures: Extraction Reagent Powd	ler (EB17)				
Chemical name	CAS No	EC No	Amount (%)	Clas	sification A 29CFR1910.1200
Sodium Lauryl Sulfate	151-21-3	205-788-1	60 to 85	Flam Oral	1. Sol. 2 H228; Acute Tox. 4 H302; Acute Tox. Inhal.
				4 H3 Dam H33	22; Skin Init. 2 H315; Eye 1 H318; STOT SE 3 Resp 5; Aquatic Tox. Chronic 3
Benzenesulfonic Acid, C10 – C13 sec-Alkyl Derivatives	4 85536-14-7	287-494-3	1.5 to 2	Acut 1C F 3 H4	e Tox. 4 H302; Skin Corr. 1314; Aquatic Tox. Chronic 112
SECTION 4. First aid measures.					
4.1 Description of first aid measures					
After inhalation	If inhaled, rem breathing is di Flush skin with contaminated	nove to fresh a fficult, give o h water. Cove clothing and s	ir. If not bre tygen. Get n r the irritated hoes. Cold y	athing, giv edical atter skin with a vater may b	e artificial respiration. If ntion immediately. an emollient. Remove e used. Wash clothing befo
After eye contact	reuse. Check for and eyes with plen	remove any c ty of water fo	ontact lenses r at least 15 r	In case o ninutes. Se	f contact, immediately flush ek medical attention if
After swallowing	Do NOT induc quantities of th tight clothing	ce vomiting u his material ar such as a colla	iless directed e swallowed ir, tie, belt or	l to do so b call a phys waistband	y medical personnel. If larg sician immediately. Loosen Never give anything by
4.2 Most important symptoms and effects, both acute and delayed	Difficulty brea Do NOT induc	athing, Skin in ce vomiting u	rson. ritation, Eye ıless directeo	irritation I to do so b	y medical personnel. If larg
4.3 Indication of any immediate medical attention and special treatment needed.	quantities of the No special treas	his material ar atment is requ	e swallowed ired	call a phys	sician immediately.
SECTION 5. Firefighting measures	5.				
5.1 Extinguishing media Suitable extinguishing agents:	SMALL FIF	RE: Use DRY Do not use v	chemical po vater jet.	wder. LAF	GE FIRE: Use water spray
5.2 Special hazards arising from the substance or mixture :	When heate	d to decompo	sition it emit	toxic fum	es of sulfur oxides, and
5.3 Advice for firefighters Protective equipment:	sodium oxid Wear approj apparatus fo	le. priate PPE for r firefighting	fire conditio	ns includin Use water	g self-contained breathing spray to cool unopened
	containers.				<i>1,</i>
SDS: EB17					
SDS: EB17					Revision nr.2
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SDS: EB17	IX on.				Revision nr.2 Dated 04/21/2019 Page n. 5 / 6
SDS: EB17	IX on.	ity LC50	Effect dose	Exposur e	Revision nr 2 Dated 04/21/2019 Page n. 5 / 6 Spectes
SDS: EB17 ENVIROLOG SECTION 12. Ecological information 12.1 Toxicity: Solium Lauryl Sulfate Aquatic toxicity: Note: Aquatic Toxicity of mixture is based on Sodium Lauryl Sulfate;	on.	ity LC50	Effect dose 10.2-22.8 mg/l	Exposur e 96 hours	Revision nr 2 Dated 04/21/2019 Page n. 5 / 6 Species Pimephales promelas
SDS: EB17 SECTION 12. Ecological informati 12.1 Toxicity: Sodium Lawyi Sulfate Aquatic oxyicy: Note Aquatic Toxicity of mixture is based on Sodium Lawyi Sulfate;	On. Aquatic toxic Acute fish tox Acute daphnii Acute dapha	ity LC50 cicity toxicity	Effect dose 10.2-22.8 mg/l 1.8 mg/l 117 mg/l	Exposur e 96 hours 48 hours 96 hours	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Species Pimephales promelas daphnia magna Pseudokirchereilla
SDS: EB17 SECTION 12. Ecological informati 12.1 Toxicity: Sodium Lawyl Sulfate Aquatic oxylery: Note Aquatic Toxicity of mixture is based on Sodium Lawyl Sulfate;	On. Aquatic toxic Acute fish tox Acute daphnia Acute algae to	ity LC50 cicity toxicity xicity	Effect dose 10.2-22.8 mg/l 11.8 mg/l 11.7 mg/l 53 mg/l	Exposur e 96 hours 48 hours 96 hours 96 hours	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Specles Pimephales promelas diphnia magna Pseudokirchnerella subcapitata Desmodesmus
SDS: EB17 SECTION 12. Ecological information 12. Toxicity: Sodium Lauryl Sulfate Aquatic toxicity: Note: Aquatic Toxicity of matthere is based on Sodium Lauryl Sulfate;	On. Aquatic toxic Acute fish tox Acute daphnii Acute algae to	ity LC50 cicity toxicity xicity	Effect dose 10.2-22.8 mg/1 1.8 mg/1 117 mg/1 53 mg/1 30-100	Exposur e 96 hours 48 hours 96 hours 96 hours 96 hours	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Species Pimephales promelas daphnia magna Pseudokirnonerilla subcapitata Desmodesmus subspiratus Desmodesmus
SDS: EB17 CONTROLOG SECTION 12. Ecological informati 13.1 Toxicity: Sodium Lawyi Sulfate Aquatic toxicity: Note Aquatic Toxicity of mixture is based on Sodium Lawyi Sulfate;	on. Aquatic toxic Acute fish tox Acute algaes to	tty LC50 icity toxicity xicity	Effect dose 10.2-22.8 mg/l 117 mg/l 53 mg/l 30-100 mg/l	Exposur e 96 hours 96 hours 96 hours	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Species Pimephales promelas daphnia magna Pseudokirchrenella subcapitata Desmodesmus subspicatus
SDS: EB17 CONTROLOG SECTION 12. Ecological informati 13.1 Toxicity: Sodium Lawyi Sulfate Aquatic toxicity: Note Aquatic Toxicity of mixture is based on Sodium Lawyi Sulfate; 12.2 Persistence and degradability :	On. Acute fish tox Acute fish tox Acute alge to Biodegradability 95.9 %	ity LC50 iicity i toxicity xiicity Result: 90 %	Effect dose 10.2-22.8 mg/l 1.8 mg/l 117 mg/l 53 mg/l 30-100 mg/l - Readily bio	Exposur e 96 hours 48 hours 96 hours 96 hours 96 hours	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Species Pimephales promelas daphnia magna Pseudokirchrenella subcapitata Desmodesmus subspicatus Desmodesmus subspicatus Lesmodesmus
SDS: EB17 SECTION 12. Ecological informati 12.1 Toxicity: Sodium Lawy Sulfate Aquatic oxycity: Note Aquatic Toxicity of mixture is based on Sodium Lawyl Sulfate; 12.2 Persistence and degradability : 12.3 Bio accumulative potential : 12.4 Mobility in sol :	On. Acute fish tox Acute fish tox Acute alges to Biodegradability 55.9 %	ity LC50 incity toxicity xicity Result: 90 % (Carp) - 72 h	Effect dose mg/1 1.8 mg/1 1.7 mg/1 53 mg/1 30-100 mg/1 - Readily bio Bioconcentu	Exposur e 96 hours 96 hours 96 hours 96 hours 96 hours adegradable	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Pimephales promelas daphnia magna Pseudokirchrenella subegriata Desmodesmus subspicatus Desmodesmus subspicatus Desmodesmus subspicatus Pseudokirchrenella subspicatus (BCF): 3.9 - 5.3
SDS: EB17 CONTROLOG SECTION 12. Ecological informati 12.1 Toxicity: Sodium Lawyi Sulfate Aquatic toxicity: Note Aquatic Toxicity of mixture is based on Sodium Lawyi Sulfate; 12.2 Persistence and degradability : 12.3 Bio accumulative potential : 12.4 Mobility in sol : 12.5 Results of PBT and vPvB assessment:	On. Acute fish too Acute fish too Acute adapting Acute adapting Acute adapting Cyptimus carpio for Not available as i	ity LC50 cicity socity	Effect dose 10.2-22.8 mg/l 117 mg/l 53 mg/l 30-100 mg/l - Readily bio Bioconcentr ety assessme	Exposur e 96 hours 96 hours 96 hours 96 hours adegradable attion factor	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Pimephales promelas daphnia magna Pseudokirchrenella subespirata Desmodesmus subspicatus L. Ratio BOD/ThBOD r(BCF): 3.9 - 5.3 ured/not conducted.
SDS: EB17	On. Acute fish too Acute fish too Acute again Acute again Acute again Cyptimus carpio fixod Not available as a No others listed No others listed	ity LC50 acity t toxicity micity Result: 90 % (Carp) - 72 h a chemical saf	Effect dose 10.2-22.8 mg/l 1.8 mg/l 117 mg/l 30-100 mg/l Bioconcentu Bioconcentu	Exposur e 96 hours 96 hours 96 hours 96 hours 96 hours 100 hours	Revision nr.2 Dated 04/21/2019 Page n. 5 / 6 Pimophales promelas daphnia magna Pseudokirchrenella subespirata Desmodesmus subspicatus Desmodesmus subspicatus . Ratio BOD/ThBOD r (BCF): 3.9 - 5.3 ured/aot conducted.
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	The se 89/686	elected protective glov 5/EEC and the standar	es have to satisfy the speci d EN 374 derived from it.	fications of EU Directive
Eye protection	Safety	glasses with side shie	lds; goggles. Use equipmorpriate enveroment char	ent for eye protection
	(US) o Eye an 1910.1	r EN 166(EU). ad face protection regulation and the second secon	lations are described by O	SHA (US) in 29 CFR ith chemicals.
SECTION 9. Physical and chemical p	ropertie	es.		
1 Information on basic physical and chemical properties	Extractio	n Reagent Powder (l	EB17)– no CAS number	
Appearance: Odor:	Solid –Po Odorless	wder, White		
Odor threshold: pH :	not applic 9.5 (1% se	able ol/water)		
Melting point/freezing point: Initial boiling point and boiling range:	No data a No data a	vailable vailable		
Flash point: Evaporation rate:	No data a No data a	vailable vailable		
Upper/lower flammability or explosive	May be co	ombustible at high ten	iperature	
Vapor pressure Vapor density	No data a No data a No data a	vailable		
Relative density Solubility(ies):	No data a Soluble in	vailable		
Partition coefficient: n-octanol/water; Auto-Ignition Temperature:	No data a No data a	vailable vailable		
Decomposition temperature: Viscosity:	No data a No data a	vailable vailable		
Explosive properties: Oxidizing Properties	No data a Not applic	vailable cable		
.2 Other information	None			
SECTION 10 Stability and reactivity				
10.1 Reactivity:	Not self-r	eactive.		
10.2 Chemical stability 10.3 Possibility of hazardous reactions :	Stable und Reaction v	er normal temperatur with strong oxidizers	es and pressures nay cause fire.	
10.4 Conditions to avoid : 10.5 Incompatible materials: 10.6 Hazardous decomposition products:	Oxidizing	agents (eg bleach).	de sulfur oridae carbon d	ioxide nitroeen oxider
2010 AMERICOUS OCCOMPOSITION PRODUCTS:	silicone O	vides.	oc, suitor oxides, carooli d	iolate, muogen oxides,
SECTION 11. Toxicological informati	on.			
Acute effects (toxicity tests):	F	Secute oral toxicity	dium lauryl sulfate - 151 LD50= 1200 mg/kg	-21-3
	F	Acute dermal toxicity	LD50= > 2000 mg/kg	rabbit
	4	Acute inhalation toxicity	LC50= 3900 mg/m3, 1hour	rat
Sensitization: N	o sensitizi	ng effects known		
Additional toxicological information:	MR (carci	nogenicity, mutageni	ity and toxicity for reprod	uction) – no CMR
ef SDS: EB17	fects.			
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			R C P	levision nr.2 lated 04/29/2019 lage n. 1 / 6
ENVIROLOGIX	Safe	ty data shee	F C P	tevision nr.2 lated 04/29/2019 /age n. 1 / 6
SECTION 1. Identification of the subst	Safe	ty data shee	F C F t company/undertaking	tevision nr.2 lated 04/29/2019 age n. 1 / 6
SECTION 1. Identification of the subst	Safe ance/mi	ty data shee	F F t company/undertaking	tevision nr.2 kated 04/29/2019 age n. 1 / 6
SECTION 1. Identification of the subst	Safet ance/mi DB 5 1 11150	ty data shee ixture and of the Dilution Buffer 1, 11665, 12495 (KR-2	r F t company/undertaking 66)	tevision nr.2 lated 04/29/2019 age n. 1 / 6
SECTION 1. Identification of the subst Trade name: Part number Part	Safet ance/mi DB 5 1 11150	ty data shee ixture and of the (Dilution Buffer), 11665, 12495 (KR-3 utory chemicals: kit or	F F company/undertaking 66)	tevision nr.2 Vated 04/29/2019 age n. 1 / 6
SECTION 1. Identification of the subst Trade name: Part number Part number 9 Alevant identifier Trade substance of the substance or mixture and uses advised against application of the substance / the preparation : 3 Details of the supplier of the safety data shee	Safet ance/mi DB 5 1 11150 Labora those s	ty data shee ixture and of the of Dilution Buffer 0, 11665, 12495 (KR-2 atory chemicals; kit co specified in product lit	t company/undertaking 66) mponent. Not to be used for erature.	tevision nr 2 ated 04/29/2019 age n. 1 / 6 g
SECTION 1. Identification of the subst Product identifier Trade name: Part number Part number Pelevant identified uses of the substance or misture and uses advised against application of the substance / the preparation : Details of the supplier of the safety data shee Manufacturer/Supplier:	Safe ance/mi DB 5 1 11150 Labors those s Envirce Portlan	ty data shee ixture and of the of Dilution Buffer b, 11665, 12495 (KR-2) atory chemicals; kit co specified in product lit of Logix, Line500 River of Logix, Line500 River	t company/undertaking 66) mponent. Not to be used fo erature. side Industrial Pkwy.	tevision nr 2 ated 04/29/2019 age n. 1 / 6 g
SECTION 1. Identification of the subst Product identifier Trade name: Part name: Part name: Part name: Part name: Delenvant identified uses of the substance or misture and uses advised against applications of the substance / the preparation : Details of the supplier: Manufacturer/Supplier:	Safet ance/mi DB 5 1 11150 Labora those 5 Enviro Portlas Phone	ty data shee http://www.commonscience.com/ ty data sheet http://www.commonscience.com/ http://wwww.commonscience.com/ http://wwwwwwwwwwwwwwwwwwwwwwwwwwwwwwwwww	F C Company/undertaking 66) mponent. Not to be used for erature. side Industrial Pkwy.	tevision nr 2 ated 04/29/2019 age n. 1 / 6 g
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SEC 15.1 S	CTION 15. Regulato safety, health and enviro	ory informat	ion.		
regula HMIS Health	ations 5 Classification (US) h hazard: 2 , Flammability de: 0	y: 1 , Physical	NFP/ Healt	A Rating (US) h hazard: 2 , Fire: 1 , Reactivity Hazard: 0	
US Fe TS	ederal Regulations SCA	a List	TSCA	A 8(b) inventory: Sodium lauryl sulfate	
CI S/	ERCLA ARA Section 302 (Extren	ig List nely Hazardou	Lister s Not li	d. isted	
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O! Europ Europ	SHA pean/International Regu ean labeling in accordanc	ilations e with EC	Not li	isted	
Directi	tives da – DSL/NDSL		This j Subst Lister	product is on the European Inventory of Existing Con ances (EINECS No. 205-788-1) d	nmercial Chemica
Canad Other	da – WHMIS		CLAS	SS D-2B: Material causing other toxic effects (TOXI a: Listed on National Inventory. Japan: Listed on Na	C). itional Inventory
			(ENC Natio	S). Korea: Listed on National Inventory (KECI). Ph nal Inventory (PICCS). Australia: Listed on AICS.	ilippines: Listed of
15.2 C	Chemical safety assessme	ent	Not c	arried out.	
SEC	CTION 16. Other inf	ormation.			
This i compi	information is true based leteness. Persons receivin	on our present ng this informa	t knowledge. I ttion must exe	However, EnviroLogix makes no representation of its rcise their independent judgment in determining the	accuracy or product's safety
and si establ EHS l	uitability for its intended lish a legally valid contra Department	use. This docu actual relations	ment shall no ship	t constitute a guarantee for any specific product feati	ires and shall not
Envir Code	oLogix Inc. Definitions:				
H228	6 + H322	Flammable	solid.	inhalad	
H315 H318	1022	Causes skin Causes serie	irritation. ous eye damaş	ge.	
H335	5	May cause r Harmful to	respiratory irri aquatic life wi	itation. ith long lasting effects.	
		Week hands	s thoroughly a		
P264 P273		Avoid relea	se to the envir	iter handling. ronment.	
P264 P273 P280 P301	+ P312	Avoid relea Wear protec IF SWALLO	se to the envir ctive gloves/ e OWED: Call a	itter handling. ronment. yep protection. a POISON CENTER/doctor/physician if you feel unv	vell.
P264 P273 P280 P301 P304 P305	+ P312 + P340 + P351 + P338	Avoid relea Wear protec IF SWALLO IF INHALE IF IN EYES	se to the envir ctive gloves/ e OWED: Call a D: Remove to S: Rinse cautio	tter handling. rye protection. POISON CENTER/doctor/physician if you feel unv fresh air and keep comfortable for breathing. usly with water for several minutes; remove contact	vell. lenses if
P264 P273 P280 P301 P304 P305 P403	+ P312 + P340 + P351 + P338 + P233	Wash hands Avoid relea Wear protec IF SWALLA IF INHALE IF IN EYES Present and Store in a w	se to the envir ctive gloves/ e OWED: Call a 2D: Remove to 3: Rinse cautio easy to do. Co rell ventilated	Itter handling. romment. ye protection. POISON CENTER/doctor/physician if you feel um 5 fresh air and keep comfortable for breathing. susly with water for several minutes, remove contact ontime rinsing. place. Keep container tightly closed	vell. lenses if
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SDS DB5 Dilution Buffer



			Revision nr.2 Dated 04/29/2019
ENVIROLOGI			Page n. 3 / 6
SECTION 6. Accidental release me	asures.		
equipment and emergency procedures:	In the case of spi a large spill, add	illed mixture wear gloves to p itional protection is recomme	revent skin contact. In the case of nded.
6.2 Environmental precautions:	Do not discharge	mixture to sewer system or v	waterways.
6.3 Methods and material for containment and cleanup:	Absorb in paper afterwards. Larg carbonate or calo	towel and discard in appropri e spills may be neutralized wi ium oxide.	ate waste. Clean with water th dilute solutions of sodium
6.4 References to other sections:	For safe handling For disposal refe	g refer to Section 7. For inforr r to Section 13	mation on PPE refer to Section 8.
SECTION 7. Handling and storage			
7.1 Precautions for safe handling:	Practice good che and clothing.	mical hygiene when handling	. Avoid contact with eyes, skin,
7.2 Conditions for safe storage, including any incompatibilities:	Store in tightly ck Prevent direct sun	osed, non-metal container, in light and heat. Store in well a	a corrosive compatible area. ired storage rooms.
7.3 Specific end use(s):	Apart from the us	es mentioned in section 1.2, n	to other specific uses are stipulated
SECTION Exposure			
8.1 Exposure limits: Components with limit values that require			
monitoring at the workplace:		EH40/2005	OSHA
8.2 Exposure Controls:	Sodium Tetraborate Decahydrate	8 Hr I WA = 5mg/m'	8 Hr 1 WA = 10 mg/m'
8.2.1Engineering controls	Facilities using t shower. Use gen below permissib	his mixture should be equippe eral or local exhaust ventilation le exposure limits.	ed with an eyewash and safety on to keep airborne concentrations
8.2.2 General protective and hygienic measures:	The usual precau	itionary measures should be a	dhered to when handling chemicals.
Eye Protection:	Safety glasses w and approved un EN 166 (EU). E in 29CFR1910.1	ith side shields, goggles. Use der appropriate government s ye and face protection regulat 33. Do not wear contact lense	equipment for eye protection tested tandards such as NIOSH (US) or ions are described by OSHA (US) as when working with chemicals
Hand Protection:	Handle with glo- removal technique with this product applicable laws a protective gloves and the standard	ves. Gloves must be inspected ue (without touching glove's + t. Dispose of contaminated gl- and good laboratory practices s have to satisfy the specificat EN 374 derived from it.	l prior to use. Use proper glove outer surface) to avoid skin contact oves after use in accordance with . Wash and dry hands. The selected ions of EU Directive 89/686/EEC
Breathing Equipment:	Appropriate resp conditions using particulate respin use respirators a government stan	iratory protection should be c risk analysis protocols. An ap ator may be used as a backup nd components tested and app dards such as NIOSH (US) or	determined according to local pproved disposable air purifying to engineering controls. Always proved under appropriate r CEN (EU).
8.2.3 Environmental exposure controls:	Contain spills, d	o not allow into environment	
SDS DB5 Dilution Buffer			
			Revision pr 2
	IX		Dated 04/29/2019
SECTION 12. Ecological information	on.		
12.1 Toxicity: Triton X-100	Fish: LC50 Pimepha Daphnia: EC50 – D	ales promelas (fathead minno aphnia – 26 mg/l – 48 hr	w) - 8.9mg/l - 96.0 hr
12.2 Persistence and degradability :	No Data Available		
12.3 Bio accumulative potential:	No Data Available		
12.4 Mobility in soil :	No Data Available		
12.5 Results of PBT and vPvB assessment:	Not available as a cl	hemical safety assessment, no	t required/not conducted.
12.6 Other adverse effects:	No Data Available		
SECTION 13. Disposal considerati	ons.		
Waste treatment methods:	Contact a license material. Dispos applicable local,	ed professional waste disposa al of surplus or waste solution state, and national laws and r	I service to dispose of this ns must be in accordance with regulations.
SECTION 14. Transport information	n.		
14.1 UN-Number DOT, ADR, ADN, IMDG, 14.2 UN proper shipping name DOT, ADR, IATA :	IATA : ADN, IMDG,	Not Hazardous for Transport	

TotalTox Aflatoxin Page 14 of 19

SECTION 9. Physical and chemical j 9. I hormation on hasic physical and chemical properties: a) Appearance: b) Odor: c) Odor Threshold: c) O	Page n. 4 / 6 properties. Clear liquid, colorless to slight yellow. None No Tata Available Xo Data Available Xo Data Available No Data Available Sable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous decompositions products should not be produced. ttor. Oral LDS0 -Rai- 1800mgkg Dermal LDS0- Rabi- 8000 mg/kg No sensiting effects known No CMR effects.
SECTION 9. Physical and chemical 1 9.1 Information on hasic physical and chemical properties: a) Appearance: b) Odor: c) Odor Threshold: c) Philing point/feezing point: c) Philing point point point c) Philing point point point point point point point point c) Philing point po	properties. Clear liquid, colorless to slight yellow. Noe Noe No Data Available No
a) Appearance: 0 b) Odor: 0 b) Odor: 0 c) Odor Threshold: 0 c) Plann built (c) (c) gascoux): 0 c) Plann built (c) (c) gascoux): 0 c) Plann built (c)	Clear liquid, colorless to slight yellow. None None No Data Available 8.6 No Data Available No farther relevant information available. y. No data available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not occ No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. ttor. Oral LD50 –Rat- 1800mg/kg Dermal LD50 -Rabi- 8000 mg/kg No sensitizing effects known No CMR effects.
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 d) pH: d) Melting point/feczing point: d) Melting point/feczing point: d) Biah point/feczing point: d) Biah point: d) Bi	 8.6 No Data Available No Data Available No tarpiciable No Data Available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not occ No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. ttor. Corll LD50 - Rat-1800mg/kg Dermal LD50. Rabbi- 8000 mg/kg No sensitizing effects known No CMR effects.
for the second sec	No Data Available. No Data Available No Data Available. No Data Available. No Data Available. No Data Available. Stable under normal temperatures and pressures. Under normal temperatures and use, hazardous decompositions produced should not be produced. ttion.
 b) Emperature rate: b) Harmability (solid, gascous): c) D) Apper/lower flammability or explosive limits: k) Vapor pressure: k) Vapor density: k) Vapor pressure: k) Vapor pressive statistical statistics: k) Vapor pressive statistics: k) Vapor stat	 No Data Available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not oct No Specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. ttor. Oral LD50 - Rat- 1800mg/kg Dermal LD50. Rabi- 8000 mg/kg No sensitizing effects known No CMR effects.
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umits: 2 1 b) Vapor pressure: 2 1 b) Vapor density 2 1 m) Relative density: 2 1 m) Solubility(ies): 4 1 o) Partiton Coefficient: n-Octanol/water: 1 p) Auto-ignition temperature: 2 1 Auto-ignition temperature:	No Uata Available No Data Available No Data Available No Data Available Fully miscible, water. No Data Available No Data Available No Data Available No Data Available No Data Available No Data Available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not oce No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. ttor.
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n) Solubility(ies:	Fully miscible, water. No Data Available No Data Available No Data Available No Data Available No Data Available No Data Available No farther relevant information available. y. No data available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not oce No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. ttor.
p) Auto-ignition temperature:	No Data Available No Data Available No Data Available No Data Available No Data Available. No further relevant information available. y. No data available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not oct No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions produces should not be produced. titon.
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b) Explosive properties: 1) Oxidizing properties:	No Data Available No further relevant information available. y. No data available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not oce No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. tton. Oral LD50 –Rat- 1800mg/kg Dermal LD50 –Rat- 1800mg/kg No sensitizing effects known No CMR effects.
9.2 Other information: SECTION 10. Stability and reactivity 10.1 Reactivity: 10.2 Chemical Stability: 10.3 Possibility of hazardous reactions: 10.4 Conditions to avoid: 10.5 Incompatible materials: 10.6 Hazardous decomposition products: SECTION 11. Toxicological informat Information on Toxicological Effects Tritos X-100 Acute toxicity: Sensitization: CMB (carcingenis): mutagenisity and toxicological information: Additional toxicological information:	No further relevant information available. y. No data available Stable under normal temperatures and pressures. Under normal conditions of storage and use, hazardous reactions will not occ No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. tton. Oral LD50 –Rat-1800mg.kg Dermal LD50 –Rat-1800mg.kg No sensitizing effects known No CMR effects.
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10 4 Conditions to avoid: 10.5 Incompatible materials: 10.6 Hazardous decomposition products: SECTION 11. Toxicological information Information on Toxicological Effects Tritos X-100 Acute toxicity: Sensitization: CMR (carcinogenity, mutagenicity and uscitry for reproduction) effects: Additional toxicological information:	No specific data No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. tition. Oral LD50 –Rat- 1800mg/kg Dermal LD50 -Rabbi- 8000 mg/kg No sensitizing effects known No CMR effects.
10.5 Incompatible materials: 10.6 Hazardous decomposition products: SECTION 11. Toxicological information Information on Toxicological Effects Triton X-100 Acute toxicity: Sensitization: CMR (carcinogenity, mutagenicity and uokicity for reproduction) effects: Additional toxicological information:	No Data Available. Under normal conditions of storage and use, hazardous decompositions products should not be produced. tition. Oral LD50 –Rat- 1800mg/kg Dermal LD50 -Rabii- 8000 mg/kg No sensitizing effects known No CMR effects.
10.6 Hazardous decomposition products: SECTION 11. Toxicological information Information on Toxicological Effects Triton X-100 Acute toxicity: Sensitization: CMR (carcinogenity, mutagenicity and molicity for reproduction) effects: Additional toxicological information:	Under normal conditions of storage and use, hazardous decompositions products should not be produced. tition. Oral LD50 – Rat- 1800mg/kg Dermal LD50- Rabbit- 8000 mg/kg No sensitizing effects known No CMR effects.
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SECTION 11. Toxicological informat Information on Toxicological Effects Triton X-100 Acute toxicity: Sensitization: CMR (carcinogenity, mutagenicity and toxicity for reproduction) effects: Additional toxicological information:	Oral LD50 -Rat- 1800mg/kg Dermal LD50- Rabbit- 8000 mg/kg No sensitizing effects known No CMR effects.
Information on Toxicological Effects Triton X-100 Acute toxicity: Sensitization: CMR (carcinogenity, mutagenicity and toxicity for reproduction) effects: Additional toxicological information:	Oral LD50 – Rat- 1800mg/kg Dermal LD50- Rabbit- 8000 mg/kg No sensitizing effects known No CMR effects.
Acute toxicity: Sensitization: CMR (carcinogenity, mutagenicity and toxicity for reproduction) effects: Additional toxicological information:	Orat L1250-etat- 18000mg/kg Dermal L1250 abbit- 8000 mg/kg No sensitizing effects known No CMR effects.
Sensitization: CMR (carcinogenity, mutagenicity and toxicity for reproduction) effects: Additional toxicological information:	No sensitizing effects known No CMR effects.
CMR (carcinogenity, mutagenicity and toxicity for reproduction) effects: Additional toxicological information:	No CMR effects.
toxicity for reproduction) effects:	No CMR effects.
Additional toxicological information:	
	No Additional Information
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SECTION 16. Other information. This information is true based on our present completeness. Persons receiving this informan and suitability for its intended use. This docu- establish a legally valid contractual relationsh EHS Department	ut knowledge. However, EnviroLogiz makes no representation of its accuracy ation must exercise their independent judgment in determining the product's ument shall not constitute a guarantee for any specific product features and s hip
EnviroLogix Inc.	
Codes:	
H302 Harmful if swallowed H315 Cause H318 Causes Serious Eye Damage H335 May c	es skin irritation H317 May cause an allergic skin reaction cause respiratory irritation H411 Toxic to Aquatic Life with Long Lasting
	Effects

SDS DB5 Dilution Buffer

14.5 Environmental hazards

14.6 Special precautions for user :

15.2 Chemical Safety Assessment

14.7 Transport in bulk according to Annex II of MARPOL73/78 and the IBC code:

SECTION 15. Regulatory information.

 14.3 Transport hazard class(es) DOT, ADR, ADN, IMDG,

 IATA):
 Not Hazardous for Transport

 14.4 Packing group (DOT, ADR, IMDG, IATA):
 Not Hazardous for Transport

No environmental hazard.

No information available.

Not carried out

None

SEC 110h to: regulatory international 15.1 Safety, health, and environmental regulations US Forderal Regulations SARA 313 Not lated US State Regulations European laternational Regulations

	Read in QuickScan: Dilution Tab on the Result Page Should Display	1:1 (this is software default)	1:A (this must be selected)	1:B (this must be selected)	
	Add Strip for	4 min.	4 min.	4 min.	
	Add Reaction Tube to Incubator Set at 22°C	Acclimate tube for 2 min [^]	Acclimate tube for 2 min^	Acclimate tube for 2 min^	
oved Matrices	Pre-mix as noted, then Transfer to Reaction Tube	<u>Pre-Mix</u> 100 μL DB5 buffer + 100 μL clarified extract in Reaction Tube	Pre-Mix400 μL Dil'n Sol'n ⁺ +100 μL clarified extractTransfer100 μL of this Pre-Mixand 100 μL DB5	Pre-Mix200 μL Dil'n Sol'n ⁺ +100 μL pre-mix extractfrom Dil ATransfer100 μL of this Pre-Mixand 100 μL DB5	
Juide for Appre	Run the Base Range Protocol First, followed by Dilution A and Dilution B Protocols, if necessary	Base Range 0 – 30 ppb	Dilution A 30 – 100 ppb	Dilution B 100 – 300 ppb (corn, sorghum)	
Summary (Clarify		Filter (corn, sorghum) <u>or</u> Centrifuge 30 sec. at 2000 x g	(corn, rice, wheat)	
	Then shake immediately		 min highest speed on shaker table or (excluding sorghum) min by hand 		
	Add to Sample Extraction Vessel (in this order)	 25g sample 1 EB17 pouch 	 75 mL water* 1mmediately shake vigorously for 10 seconds by hand OR (excluding wheat) 50g sample 	 2. 2 EB17 pouches 3. 150 mL water* 4. Immediately shake vigorously for 10 seconds by hand 	
	Approved Matrix	Сот	(MG1) Sorghum (MG4) Brown Rice (MG7) Wheat	(MG8) (70-80% through 20-mesh)	

Notes:

*Use distilled, deionized, or flat (non-carbonated) bottled water †Dilution Solution = Mix 1 x EB17 pouch with 150 mL water

^The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F)

Rev 5. Effective 1/29/2024 IFS CO: 23-0237

	Read in QuickScan: Dilution Tab Should Display	1:1 (this is software default)	1:A (this must be selected)	1:B (this must be selected)	1:1 (this is software default)
	Add Strip for	5 min.	5 min.	5 min.	4 min.
	Add Reaction Tube to Incubator Set at 22°C	Acclimate tube for 2 min^	Acclimate tube for 2 min^	Acclimate tube for 2 min [^]	Acclimate tube for 2 min^
d Matrices (cont.)	Pre-mix as noted, then Transfer to Reaction Tube	<u>Pre-Mix</u> 100 μL DB5 buffer + 100 μL clarified extract in Reaction Tube	Pre-Mix400 μL Dil'n Sol'n [†] +100 μL clarified extractTransfer100 μL of this Pre-Mixand 100 μL DB5	Pre-Mix200 μL Dil'n Sol'nt +100 μL pre-mix extract from Dil ATransfer100 μL of this Pre-Mixand 100 μL DB5	<u>Pre-Mix</u> 100 μL DB5 buffer + 200 μL extract in Reaction Tube
e for Approved	Run the Base Range, then Dilution A and Dilution B Protocols	Base Range 0 – 30 ppb	Dilution A 30 – 100 ppb	Dilution B 100 – 300 ppb (<i>barley</i>)	Base Range 0 – 30 ppb
nmary Guid	Clarify		Centrifuge 30 sec. at 2000 x g		<u>Immediately</u> Filter <u>or</u> Centrifuge 1 min. at 2000 x g
Sun	Then shake immediately		 min highest speed on shaker table 		 min highest speed on shaker table <u>or</u> hand
	Add to Sample Extraction Vessel (in this order)		 25g sample 1 EB17 pouch 75 mL water* 1 Inmediately shake vigorously for 10 seconds by 	hand	 10g sample 1 EB17 pouch 60 mL water* Immediately shake vigorously for 10 seconds by hand
	Approved Matrix		Barley (MG18) Oats (MG22)		Masa flour (MG5), Corn flour (MG6)

Notes:

*Use distilled, deionized, or flat (non-carbonated) bottled water †Dilution Solution = Mix 1 x EB17 pouch with 150 mL water

^The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F)

Rev 4. Effective 12/1/2023 IFS CO: 23-0228

	Read in QuickScan: Dilution Tab Should Display	1:1 (this is software default)	1:A (this must be	selected)	1:B (this must be selected)	1:1 (this is software default)	
	Add Strip for			5 min.			
ed Matrices (cont.)	Add Reaction Tube to Incubator Set at 22°C			Acclimate tube for 2 min^			
	Pre-mix as noted, then Transfer to Reaction Tube	<u>Pre-Mix</u> 100 μL DB5 buffer + 100 μL clarified extract in Reaction Tube	Pre-Mix400 μL 50% ethanol + 100μL clarified extractTransfer100 μL of this Pre-Mixand 100 μL DB5	Pre-Mix300 μL 50% ethanol + 100μL clarified extractTransfer100 μL of this Pre-Mixand 100 μL DB5	Pre-Mix200 μL 50% ethanol +100 μL pre-mix extractfrom Dil ATransfer100 μL of this Pre-Mixand 100 μL DB5	<u>Pre-Mix</u> 100 μL DB5 buffer + 100 μL clarified extract in Reaction Tube	
le for Approve	Run the Base Range, then Dilution A and Dilution B Protocols	Base Range 0 – 30 ppb	Dilution A 30 – 100 ppb <i>(DDGS,</i> <i>corn germ,</i> <i>cottonseed, rye)</i>	Dilution A 30 – 100 ppb (Corn gluten meal)	Dilution B 100 – 300 ppb <i>(DDGS,</i> corn germ, rye)	Base Range 0 – 20 ppb	
nmary Guid	Clarify			Centrifuge 1 min. at 2000 x g <u>or</u>	Filter (DDGS only*)		
Sun	Then shake immediately	1 min highest speed on shaker table <u>or</u> (<i>excluding rye</i>) 2 min by hand					
	Add to Sample Extraction Vessel (in this order)		 25g 23mple 100 mL 50% ethanol 	<i>OR</i> 1. 50g 2. 200 mL	50% ethanol	 10 g sample 40 mL 50% ethanol 	
	Approved Matrix		DDGS (MG2) Corn Germ (MG10)	Corn Gluten Meal (MG12) Cotton-seed (MG17)	Rye (MG21)	Corn Fermented Protein (MG26)	

<u>Notes</u>: $^{\wedge}$ The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F) *Only DDGS can be clarified via the filter method for ethanol-extraction matrices.

Rev 4. Effective 12/1/2023 IFS CO: 23-0228

		Sun	nmary Guid	le for Approve	d Matrices (cont.)			
				Run the Base Range, then		Add Reaction		Read in OuickScan:
	Add to Sample	Then also		Dilution A and	Pre-mix as noted, then	Tube to	Add	Dilution Tab
Approveu Matrix	(in this order)	immediately	Clarify	Protocols	Reaction Tube	Set at 22°C	for	Display
Sovhean	1. 25g sample 2. 50 mL 50% ethanol	1 min highest speed on	Centri filoe	Base Range 0 – 30 ppb	Pre-Mix 100 μL DB5 buffer + 100 μL clarified extract in Reaction Tube	Acclimate tube for 2 min^	5 min.	1:1 (this is software default)
Meal (MG15)	<i>OR</i> 1. 50g sample 2. 100 mL 50% ethanol	shaker table <u>or</u> hand	1 min. at 2000 x g	Dilution A 30 – 100 ppb	Pre-Mix400 μL 50% ethanol + 100μL clarified extract100 μL of this Pre-Mixand 100 μL DB5	Acclimate tube for 2 min^	5 min.	1:A (this must be selected)
Corn High Sensi- tivity (MG9)	 25g sample 40 mL 50% ethanol OR 50% ethanol 80 mL 50% ethanol 	 min highest speed on shaker table <u>or</u> 2 min by hand 	Centrifuge 1 min. at 2000 x g	Base Range 0 – 10 ppb	<u>Pre-Mix</u> 100 μL DB5 buffer + 100 μL clarified extract in Reaction Tube	Acclimate tube for 2 min^	5 min.	1:1 (this is software default)
Corn	1. 25g sample	2 min highest	Settle 30 sec.	Base Range 0 – 50 ppb	<u>Pre-Mix</u> 175 μL DB5 buffer + 25 μL clarified extract in Reaction Tube	Acclimate tube for 2 min^	5 min.	1:1 (this is software default)
Feed (MG13)	 35 mL 84% acetonitrile* 	speed on shaker table	<u>or</u> Centrifuge 30 sec. at 2000 x g	Dilution A 50 – 200 ppb	Pre-Mix500 μL 84% acetonitrile +100 μL clar. extract25 μL of this Pre-Mixand 175 μL DB5	Acclimate tube for 2 min^	5 min.	1:A (this must be selected)
			ر	•				

<u>Notes</u>: *Acetonitrile may leak when extracting; refer to page 3 for preventative measures. ^The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F)

	Read in QuickScan: Dilution Tab Should Display	1:1 (this is software default)
	Add Strip for	4 min.
Summary Guide for Approved Matrices (cont.)	Add Reaction Tube to Incubator Set at 22°C	Acclimate tube for 2 min^
	Pre-mix as noted, then Transfer to Reaction Tube	Pre-Mix 300 μL DB5 buffer + 100 μL clarified extract in Reaction Tube
	Run the Base Range, then Dilution A and Dilution B Protocols	Base Range 0 – 30 ppb
	Clarify	Centrifuge 1 min. at 2000 x g, pour supernatant into separate vessel
	Then shake immediately	 min highest speed on shaker table or 2 min by hand
	Add to Sample Extraction Vessel (in this order)	 0.8 mL/g sample salt water to sample Mix well, stir slowly 2 mL/g sample 80% Ethanol
	Approved Matrix	Peanut Seed (MG27) Peanut Oil (MG28)

[^]The tube acclimation step is only required if the temperature of the testing environment is unknown or outside of 20 - 24°C (68 - 75°F)